

Molecular Detection of Colorectal Cancer

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Molecular Detection of Colorectal Cancer

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Definition

Drug-specific therapeutic approaches for colorectal cancer (CRC) have contributed to a significant improvement in the health status of patients. However, a great need to improve personalization of treatments based on genetic and epigenetic tumor profiles to maximize quality and efficacy while limiting cytotoxicity remains. Currently, CEA and CA 19-9 are the only validated blood biomarkers in clinical practice. For this reason, laboratories are trying to identify new specific prognostic and, more importantly, predictive biomarkers for CRC patient profiles. Thus, the unique landscape of personalized biomarker data should have a clinical impact on CRC treatment strategies and molecular genetic screening tests should become the standard method for CRC diagnosis, as well as detection of disease progression.

1. Introduction

Colorectal cancer (CRC) represents the second leading cause of cancer-related death worldwide ^[1], with an annual incidence of nearly two million cases ^[2] (**Figure 1**). Moreover, the CRC incidence is increasing in low- and middle-income countries ^[1]. The disease results from the accumulation of multiple genetic and epigenetic modifications that lead to the transformation of colonic epithelial cells into invasive and aggressive adenocarcinomas ^{[3][4]}. The lack of and inadequate response to numerous mono-target therapies in cancer treatment emphasizes the need for personalized diagnostic and therapeutic approaches for effective strategies that target not only tumor cells, but more importantly the multicellular tumor microenvironment to improve patient outcomes. Nevertheless, one of the most important keys to the successful treatment of this malignant tumor and patient survival is not only the early diagnosis of the disease but also the control of tumor dissemination and progression ^[5]. Thus, the 5-year survival rate for patients with early diagnosis is approximately 90%. In contrast, the survival rate for patients with regional lymph node metastasis is about 70%, and for those with distant metastases is only 13% ^{[6][7]}.

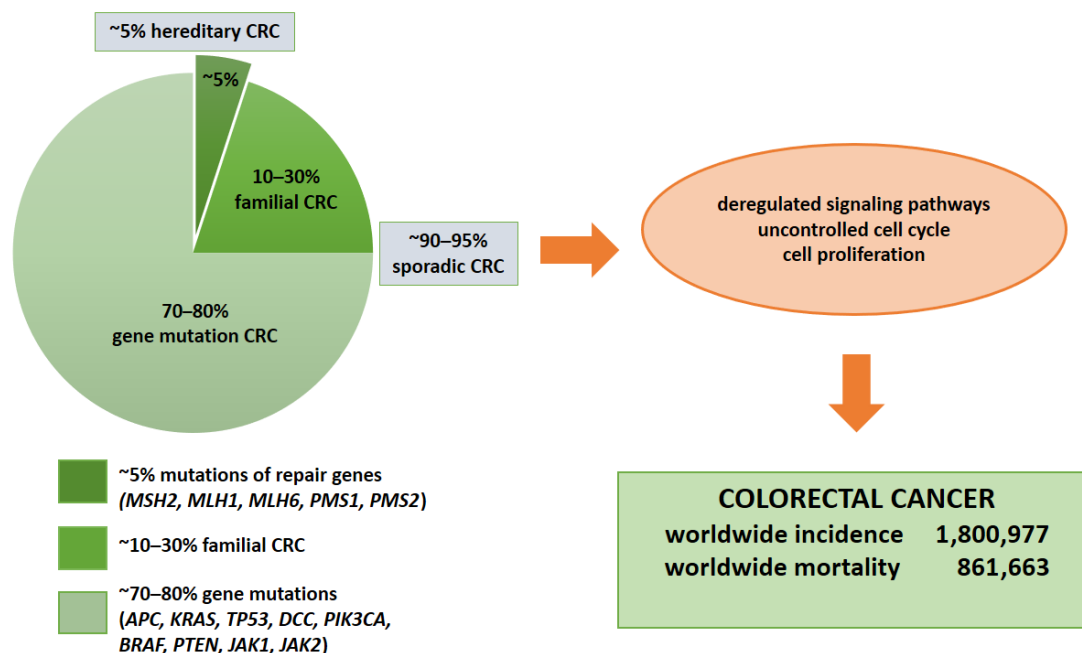


Figure 1. Molecular basis of CRC. Colorectal cancer is based on gene mutations, familial or hereditary CRC. The indication of total numbers refers to Global Cancer Statistics 2018 [3]. For worldwide incidence and mortality, colorectal cancer cases from 185 countries in 2018 were summed.

Other important keys to improving CRC therapies include improvements in surgical modalities and adjuvant chemotherapy, which has increased cure rates for early-stage disease. However, a significant proportion of patients will unfortunately develop recurrence or advanced disease. Nevertheless, the efficacy of chemotherapy for recurrence and advanced stages of CRC has improved significantly over the past decade. Previously, the historical drug 5-fluorouracil was the only chemotherapeutic agent used. With the addition of other chemotherapeutic agents such as capecitabine, irinotecan, oxaliplatin, bevacizumab, cetuximab, panitumumab, vemurafenib, and dabrafenib, the median survival of patients with oligometastatic CRC has improved significantly from less than one year to the current standard of nearly two years [8]. However, many side effects of systemic therapy, such as toxicity, can lead to fatal complications and significantly affect patients' quality of life. In parallel, a plethora of biologically active compounds are being tested *in vitro* and *in vivo*, and promising hits/leads compounds are being identified for development as adjuncts to therapy [9][10]. Thus, there is an urgent need for crucial biomarkers to select optimal drugs individually or in combination for an individual patient. The application of personalized therapy based on DNA testing could help clinicians provide the most effective chemotherapeutic agents and dose modifications for each patient. However, some of the current findings are controversial, and the evidence is conflicting [11]. The current trend is to achieve successful personalized therapeutic approaches based on monitoring disease-specific biomarker(s). Finally, data in this respect are scarce, and studies involving personalized testing versus treatment are needed.

2. Etiology of Colorectal Cancer

The CRC-related genes and pathways often overlap with other solid tumors, such as breast and prostate cancer. In approximately 70-90% of patients, CRC develops sporadically due to point mutations in the *APC*, *KRAS*, *TP53*, and *DCC* genes.

In approximately 1-5% of cases, it is a consequence of hereditary polypoid and non-polypoid syndrome and 10-30% of patients have a familial CRC [12] (Figure 1). It is important to note that 1-2% of CRC are associated with chronic inflammatory diseases such as ulcerative colitis and Crohn's disease. The risk increases with longer duration of ongoing inflammation [13], which is also caused by dysbiosis in the gut [14] and inappropriate dietary patterns and deteriorating life-style conditions [15]. Chromosomal instability

(CIN) as a major molecular pathway of malignant transformation mainly affects genes such as *APC*, *KRAS*, *PIK3CA* and *TP53* [16]. In addition, the adenoma–carcinoma sequence offers potential for screening and surveillance; for example, expression of connexin 43 in colonic adenomas is associated with high-grade dysplasia and colonic mucosa surrounding adenomas [17]. *APC* mutations lead to nuclear beta-catenin translocations and transcription of genes involved in carcinogenesis and invasion processes. *KRAS* and *PIK3CA* mutations lead to continuous activation of mitogen-activated protein kinase (MAPK) signaling pathways, which in turn increases cell proliferation, while *TP53* mutations lead to loss of p53 function and uncontrolled cell cycle [16]. Finally, epigenetic instability (CIMP) is associated with hypermethylation of the promoter of oncogenes and loss of expression of the corresponding proteins [18].

3. The impact of Genetic Alterations on Disease Outcome

The most common mutations, chromosomal alterations and translocations affect critical wingless-related integration sites (WNT), MAPK/phosphatidylinositol 3-kinase (PI3K) and transforming growth factor β (TGF- β) signaling pathways and intracellular protein functions such as p53, as well as cell cycle regulation [19]. The WNT signaling pathway, which is a critical mediator of tissue homeostasis and repair, is frequently co-opted during tumor development. Almost all colorectal cancers demonstrate hyperactivation of the WNT pathway, which is considered to be the initiating and driving event in many cases [20]. *APC* gene mutations represent the most significant genetic alteration associated with the WNT signaling pathway, which regulates stem cell differentiation and cell growth. Nevertheless, due to their high CRC frequency and the number of different mutations identified within the gene, they are not a good predictor of disease progression [21]. Increased β -catenin expression associated with the WNT signaling pathway has also been found to be an unreliable marker for disease prognosis. In contrast, overexpression of the *c-MYC* gene triggered by activation of the WNT signaling pathway represents a good predictor of metastasis and disease progression [22][23]. *KRAS*, *BRAF*, and *PIK3CA* mutations are common and associated with the MAPK/PI3K signaling pathways.

Furthermore, mutations of the *KRAS* gene in exon 2, codon 13 are associated with poor prognosis and a low survival rate, while mutations in exon 2 and codon 12 are associated with tumor progression and metastasis [24][25]. Recently, AMG 510, the first *KRAS* G12C inhibitor, has entered into the clinical development after promising preclinical results [26]. These are intriguing efforts that could overcome the notion that *KRAS* is in principle "undruggable" as a therapeutic target and may contribute to the development of effective drugs for targeting traditionally difficult signaling pathways in the clinical setting [27]. *BRAF* gene mutations are associated with poor prognosis and survival [28][29][30].

The associations between disease outcome or survival and *PIK3CA* mutations have not yet been established. Nevertheless, there is evidence that these mutations, in combination with the *KRAS* gene mutations, are associated with poor outcome [31]. Moreover, CRC patients with multiple *PIK3CA* mutations, e.g., a combination of mutations in exons 9 and 20, have a worse prognosis than patients with only one of these mutations [32]. Protein phosphatase and tensin homolog (PTEN) adversely affects the PI3K signaling pathway and CRC in which loss of the *PTEN* gene has been associated with poor prognosis [33]. In CRC patients, alterations in the TGF- β signaling pathway are associated with CIN [34]. Chromosome 18q carries the tumor suppressor genes *SMAD2* and *SMAD4*, whose encoded proteins are functionally associated with apoptosis and cell cycle regulation [35][36]. Similarly, they play a role in tumor cell migration by regulating the activity of proteins such as matrix metalloproteinase 9 (MMP9) [37]. A significant association between the loss of chromosome 18q and poor prognosis and survival has not been found [35][36]. In CRC, loss of the *17q-TP53* gene, which encodes the tumor suppressor protein p53 that regulates the cell cycle, is quite common. Without it, cells proliferate uncontrollably and the tumor progresses [38]. Janus kinases, JAK1 and JAK2, are associated with cytokine receptors [39][40], and cytokine binding leads to their activation and phosphorylation. Subsequently, Janus kinases phosphorylate signal transducer and activator of transcription (STAT) proteins, leading to their translocation to the nucleus and transcription of their target genes [39][40]. There is evidence of *JAK1* and *JAK2* gene mutations that inhibit the function of the corresponding JAK1 and JAK2 proteins. The *JAK1* frameshift mutations (positions 142/143, 430/431, and

860/861) have been described as the consequence of insertion/deletion of a nucleotide [40]. The V617E mutation results in a *JAK2* loss-of-function mutation [39]. These mutations were found in tumors with high MSI resulting from dysfunctional DNA repair during replication, known as mismatched repair [41]. Indeed, they were associated with tumor resistance to treatment targeting programmed cell death protein 1 (PD-1) [41]. The determination of molecular alterations at the DNA level, particularly derived from tumor-specific liquid components such as circulating tumor DNA (ctDNA), exosomes, and circulating tumor cells (CTC), can improve the prediction of disease progression and help in adjustment of therapy for each patient in the context of personalized medicine.

4. Liquid Biopsy

Much has been learned about the molecular background of the development and progression of CRC, which can help to tailor therapy for each individual patient and improve their survival prognosis. However, early CRC still relies on biomarkers from readily available biological materials (**Table 1**). Currently, there are only two validated protein-based blood biomarkers used in routine clinical practice: carcinoembryonic antigen (CEA) and carbohydrate antigen 19-9 (CA 19-9). CEA is an embryo-specific glycoprotein that can also be found in CRC. In clinical practice, it is used to monitor tumor progression after diagnosis [42]. However, it shows insufficient sensitivity and specificity because it is hereditarily determined and in the case of recessive homozygote, the levels of CA 19-9 would not be elevated (in approximately 15% of individuals) [43][44][45]. During the search for new biomarkers that could replace the old ones, a promising non-invasive and repeatable procedure called “liquid biopsy” has been developed for different body fluids (blood, saliva and urine). Liquid biopsy from peripheral blood is used for diagnostic screening as well as for determining response to therapy and evaluating the outcome of the disease [46]. Peripheral blood may contain CTC, ctDNA, and exosomes (vesicular structures containing proteins and RNA molecules that can be released into circulation by various cells, including tumor cells) [47]. This could allow the molecular profile of the disease, the degree of affected tissue, and the response to therapy to be determined in a non-invasive manner. The founder and establisher of new principles and methods of healing, Leroy Hood, has relentlessly emphasized that in the new era of personalized approaches undertaken while assessing different conditions and diseases, “the blood becomes a window through which we observe what is happening in the body” [48]. The same idea has been accepted and maintained by other biomedical disciplines, from genetics to personalized nutrition [49]. Future molecular profiling, ideally assessed and monitored by liquid biopsy, could further personalize decision-making in the adjuvant setting of CRC [50][51].

However, the liquid biopsy results have to be combined and evaluated with the pathological findings of the tissue before final validation of the proposed approach. The existing testing landscape presents additional challenges in the application of liquid biopsy in clinical practice, and consideration needs to be given to how the pathologist should be involved in the interpretation of liquid biopsy data in the context of the patient’s cancer diagnosis and stage assessment [52].

Table 1. Structure, signification and implication related to relevant biomarkers of CRC.			
Biomarker (Denotation)	Structure	Experience/Implication	Reference
CEA (carcinoembryonic antigen)	glycoprotein	Validated blood biomarker in clinical practice. Not recommended as a sole CRC screening test. Preoperative CEA > 5 mg/mL may correlate with poorer CRC prognosis. Used as a postoperative serum test and for monitoring every 3 months during active CRC treatment. Diagnostic sensitivity 54.5%; specificity 98.4%.	Locker et al.,2006 [42] Wu et al., 2020 [53]

Biomarker (Denotation)	Structure	Experience/Implication	Reference
CA 19-9 (carbohydrate antigen)	glycoprotein	Validated blood biomarker in clinical practice. Not recommended as a sole screening or monitoring CRC marker. Used as a supplementary progress monitoring during pancreatic cancer treatment every 1–3 months. Individual values for each patient. Diagnostic sensitivity 64.4%; specificity 96.8%.	Locker et al., 2006 [42] Wu et al., 2020 [53]
CTC (circulating tumor cells)	tumor cells	Epithelial marker in peripheral blood via automatic detection system. Detected in various cancers. 1–10 CTCs per ml of blood were found in patients with metastases but rarely in healthy individuals. Poor prognosis for CRC patients with ≥ 5 CTC per 7.5 ml of blood. CTC count ≥ 5 was an independent prognostic factor for distant metastasis (hazard ratio = 7.5, 95% CI: 1.6 to 34.7, $p = 0.01$). Diagnostic sensitivity 62.7%; specificity 82.0%.	Dominguez-Vigil et al., 2018 [47] Tsai et al., 2016 [54]
ctDNA (circulating tumor DNA)	small DNA fragments released by tumor cells	Tumor mutation search in peripheral blood, plasma and serum. Patients with 100 g tumor burden released 3.3% of ctDNA into circulation. In CRC, ctDNA is more sensitive than CEA. KRAS mutations were detected with 87.2% sensitivity and 99.2% specificity.	Osumi et al., 2020 [55] Said et al., 2020 [56] Dominguez-Vigil et al., 2018 [47]
exosomes (nanovesicles)	vesicular structures released by different cell types, including tumors	Tumor miRNA molecules in biological fluid such as blood and urine. Associated with several types of CRC. Each tumor is characterized by a specific protein profile. Positive correlation of exosome miRNA and proteins with the stage of tumor progression.	Dominguez-Vigil et al., 2018 [47] Wang et al., 2016 [57]

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