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CASE REPORT

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Unusual presentation of plasmablastic lymphoma involving ovarian mature cystic teratoma: a case report

Ita Hadžisejdić^{1*} , Emina Babarović¹, Luka Vranić², Antica Duletić Načinović³, Ksenija Lučin¹, Maja Krašević¹ and Nives Jonjić¹

Abstract

Background: Plasmablastic lymphoma (PBL) is relatively new clinical entity described as a distinct subtype of diffuse large B-cell lymphoma (DLBCL). It is characterized by its aggressive nature and proliferation of large neoplastic cells resembling immunoblasts including cells with more obvious plasmacytic differentiation. In this case report, we describe an unexpected finding of PBL associated with a mature cystic teratoma of the ovary in a young immune competent woman.

Case presentation: A 19-year old woman was admitted to the hospital with generalized lymphadenopathy, a pelvic tumor mass measuring 35 × 30 cm and a 4 cm lump in her right breast. She underwent a right salpingo-oophorectomy, lymphadenectomy, splenectomy, omentectomy, and a right breast lumpectomy. On macroscopic examination the right ovary was replaced by a thick-walled multilocular cystic tumor. Upon incision, the cysts were filled with thick, greasy sebaceous material and hair and there were several solid nodules within the cyst walls. Histological examination revealed a mature cystic teratoma and malignant non-Hodgkin lymphoma (NHL) within the solid nodules. Tumor tissue from the right breast, spleen and lymph nodes, all had the same histological, NHL morphology. After extensive immunostaining, a diagnosis of PBL was made. Following surgery, the patient was treated with different chemotherapy regimens, without any significant regression of the disease, and died of multiple organ failure.

Conclusions: Primary NHL of the ovary is relatively rare occurrence while secondary involvement by lymphoma is much more common. PBL is a rare lymphoma, primarily reported in the jaw and oral mucosa, but also documented in extra-oral sites. To the best of our knowledge, this is the first case described in a mature ovarian cystic teratoma. Although the patient was HIV-negative and immune competent, she had progressive disease and died despite aggressive chemotherapy 11 months after the initial diagnosis.

Keywords: Plasmablastic lymphoma, Ovarian mature cystic teratoma, Immune competent patient

Background

Plasmablastic lymphoma (PBL) is a rare and aggressive variant of NHL with diffuse proliferation of large neoplastic cells with cytomorphological features that resemble B immunoblasts, but with the immunophenotype of the plasma cells [1]. In the most cases, this neoplasm occurs in the oral cavity of HIV-positive patients. Other immunodeficiency states such as iatrogenic immune suppression in treatment of autoimmune diseases or chemotherapy are

also associated with this diagnosis [1, 2]. It is primarily an adult disease that occurs most frequently around 50 years of age, affecting men more often than women. Although, the majority of patients are adults, it has also been reported in the pediatric age group [3]. Involvement of extra-oral sites, such as the orbit, sinonasal cavity, gastrointestinal tract, soft tissues, liver, retroperitoneal region and bone can also be found, especially in PBLs that are not associated with HIV infection [3]. The PBL cells are usually positive for a plasma cell phenotype including CD138, CD38, CD79a, IRF4/MUM1, cytoplasmic immunoglobulins (most frequently IgG) and either kappa or lambda light chain with a high Ki-67 proliferation index (usually ≥80%) [2, 3].

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Approximately 70% of cases express EBV-encoded RNA (EBER), which is the most sensitive methodology for detecting EBV infection within the malignant cells [2]. Both HIV-positive and HIV-negative patients are at advanced clinical stage (Ann-Arbour III or IV) on presentation. This rare clinical entity represents a diagnostic challenge due to its specific morphological and immunohistochemical resemblance to lymphomas with plasmablastic differentiation, more precisely plasmablastic or anaplastic multiple myeloma. Furthermore, with its aggressive clinical course and almost 100% rate of death within the first year of diagnosis, it also remains a therapeutic challenge. This report presents a very unusual case of PBL in an immune competent, HIV-negative, 19-year old woman, that was detected within the walls of an ovarian cystic teratoma occurring as a part of systemic disease with involvement of the breast, spleen and lymph nodes.

Case presentation

A 19-year old female was referred to the Department of Internal Medicine for workup of anemia. She complained of having fever (up to 38 °C), night sweats, dyspnea, dry cough, and fatigue. Symptoms had been present for 4 weeks and had not improved after a course of antibiotics prescribed by her family physician. On physical examination, a palpable painless mass was noticed in her right abdomen, with enlarged inguinal lymph nodes as well as a palpable lump in the right breast with ipsilateral axillary lymphadenopathy. Abdominal ultrasound (US) examination revealed a multiloculated cystic mass with solid areas measuring 35 × 30 cm in her pelvis. The US examination also showed an enlarged spleen with enlarged para-aortal lymph nodes. Subsequently, a thoracic, abdominal and pelvic multi-slice computed tomography scan (MSCT) was performed and showed a well circumscribed, solid nodule, 4 cm in diameter, in the upper outer quadrant of right breast with enlarged ipsilateral axillary lymph nodes, along with generalized abdominal lymphadenopathy, an enlarged spleen with diffuse multiple nodal masses and a complex right pelvic tumor mass with amorphous calcification similar to tooth formation. Based on the MSCT scan, a tentative diagnosis of an immature teratoma was made. The patient proceeded to surgery for a unilateral right salpingo-oophorectomy, lymphadenectomy, splenectomy, omentectomy and right breast lumpectomy. Intraoperative peritoneal washing revealed a poorly differentiated malignant tumor on cytology.

On macroscopic examination, the right ovary was replaced by a cystic mass with a glistening surface, measuring 17 cm in diameter (Fig. 1a). On incision, the cyst was partly filled with fatty material and hair. The wall of the cyst was thickened with solid, grayish nodules. The spleen was diffusely infiltrated with grayish nodes as well, measuring up to 4.5 cm in diameter (Fig. 1b). There was a lobulated, well circumscribed solid, grayish

nodule identified on lumpectomy of the breast. On histological examination, the cavity of the cyst was lined by skin with dermal appendages as well as respiratory type of epithelium, smooth muscle, mature adipose and bone tissue. This morphology was in agreement with a diagnosis of mature cystic teratoma of the ovary. Solid nodules within cyst walls were tumor tissue, with a diffuse growth pattern, composed of large polymorphic cells with vesicular nuclei, a high nuclear-cytoplasmic ratio, and one or more prominent nucleoli. Some cells had eccentrically positioned nuclei with a cytoplasmic halo resembling plasmablasts. The tumor tissue from the right breast and spleen had the same histological appearance. An extensive immunohistochemical evaluation was performed. Tumor cells were negative for panCK, AFP, PLAP, CD117, D2-40, CD30, MyoD1, HMB45, desmin and CD99, but positive for LCA, EMA and vimentin. Therefore, a diagnosis of NHL was made. Subsequent immunohistochemistry showed tumor cells to be CD79a, PAX5, MUM-1, CD38, CD138 positive and CD20, CD10, Bcl-6, ALK, cyclin D1, CD56 negative. Ki-67 was high, around 80% and EBER in situ hybridization was negative (Fig. 2). According to the morphological and immunohistochemical findings a diagnosis of PBL was made. Based on the clinical findings, the patient was staged, Ann-Arbour IV PBL. Laboratory data showed anemia, normal platelet and leucocyte counts with elevated lactate dehydrogenase (LDH), aspartate aminotransferase (AST) and C-reactive protein (CRP). Serum alpha-1-fetoprotein and human chorionic gonadotropin were normal while carbohydrate antigen 125 was increased. Urine analysis and renal function were within normal limits. Serum immunoglobulin levels were within normal limits, without evidence of monoclonal gammopathy and serology was negative for HIV, HBV and HCV. The patient was transferred to the Hematology ward and treated with dose-adjusted etoposide, prednisone, vincristine, cyclophosphamide, and doxorubicin (DA-EPOCH) as well as intra-theal EPOCH with each cycle [2, 4]. After 4 cycles of chemotherapy, the patient subjectively felt better however evaluation with positron emission tomography and computed tomography scans (PET-CT) showed high activity of disease. An autologous peripheral blood stem cell transplantation was planned and 1 cycle of salvage chemotherapy was given according to DHAP protocol (cisplatin, cytarabine and dexamethasone). Unfortunately, her underlying disease continued to progress and 4 cycles of alternating modified CODOX-M and IVAC chemotherapy (cyclophosphamide, vincristine, doxorubicin, high-dose methotrexate/ifosfamide, etoposide, and high-dose cytarabine) were given. Autologous stem cell transplantation could no longer be performed due to patient's clinical deterioration. After the fourth cycle of

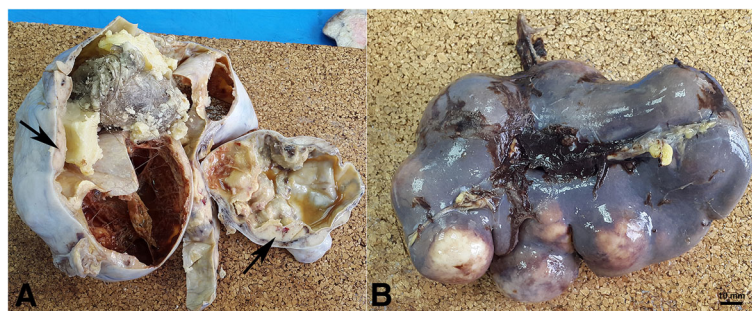


Fig. 1 Macroscopic image of multiloculated ovarian cyst and spleen. **a** The right ovary was replaced by a cystic tumor (mature teratoma) with multiple solid nodules of PBL within the cyst's walls (arrows). **b** Spleen with multiple PBL nodules

chemotherapy, PET-CT scan was performed again, and revealed profound disease progression. The patient also had developed treatment complications, such as secondary anemia, thrombocytopenia, sepsis, enterocolitis, ileus, pleural effusions and ascites. Due to the worsening of the patient's physical condition, her goals of care were changed and only supportive therapy was given. The patient died of multiple organ failure approximately 11 months after initial diagnosis (Additional file 1).

Discussion

PBL was originally reported exclusively in the jaw and oral mucosa of male, predominantly HIV-positive, patients [5]. It may be present in patients who are HIV-negative, but otherwise immune compromised such as patients with solid organ or bone marrow transplantation or those being treated for autoimmune disorders [6]. Cases of PBL occurring in elderly, immune competent patients have been documented and increasingly described in extra-oral locations [1, 6, 7]. Extra-oral PBL has been reported in various locations; however to the best of our knowledge, PBL was never described within ovarian cystic teratoma. This case is unusual as it presents the first described case of PBL in a 19-year old, immune competent patient, within the walls of a mature ovarian cystic teratoma, with concurrent involvement of spleen, lymph nodes and breast tissue. Primary NHL of the ovary is a relatively rare occurrence while secondary involvement is much more common [8]. Descriptions of NHL arising in mature cystic teratoma are even more rare. There have only been a few reports describing DLBC and follicular lymphoma arising in a mature cystic teratoma [9–11]. Primary ovarian lymphoma accounts for 0.5% of extranodal NHL and 1.5% of primary ovarian cancers [12]. The existence of primary ovarian NHL is a question of debate by some authors because there is no pre-existing lymphoid tissue in the ovaries for lymphoma to develop. However, some studies have identified small number of lymphocytes around blood vessels at the hilum and within epithelial structures of teratomas, supporting the hypothesis for possible

development of primary ovarian lymphoma [13, 14] from this lymphoid tissue. Also, due to varying definitions, the existence of primary ovarian lymphoma remains debatable. Fox et al. proposed the following diagnostic criteria for a primary ovarian lymphoma: 1. The lymphoma should be confined to the ovary or at the maximum, to the adjacent group of lymph nodes draining the organ involved or if there has been direct infiltration of adjacent structures while full investigation fails to reveal evidence of lymphoma elsewhere; 2. The peripheral blood and bone marrow should not contain any abnormal cells; 3. If further lymphomatous lesions occur at sites remote from the ovary, then at least several months should have elapsed between the appearance of the ovarian and extra-ovarian lesions [15]. According to this definition, our patient's PBL was disseminated, secondary disease with concurrent involvement of the ovarian mature cystic teratoma. Also, our patient presented with B symptoms as a main complaint, which is more suggestive of secondary involvement by PBL because the majority of patients with primary ovarian lymphomas present with abdominal or pelvic pain as their main complaint. Possible origin from a teratoma with differentiation along hematopoietic cell lines could be considered, but it is less likely and difficult or impossible to prove.

EBV plays an important role in the tumorigenesis of PBL with expression of EBER by in situ hybridization (ISH) in 82% and 46% of the HIV-positive and HIV-negative cases, respectively [3]. The presence of plasmablasts is noted in reactive processes, associated with viral infections, but nevertheless pathogenesis of PBL is poorly understood.

In addition to EBV infection, recent studies have identified the presence of *MYC* gene rearrangements as important pathogenic mechanisms [16, 17]. Using ISH, Valera et al., examined a larger population of PBL patients and found *MYC* rearrangement in 49% of the cases [18]. In PBL, *MYC* deregulation, mediated by translocation or amplification allows *MYC* to overcome the regulatory effects of BCL-6 or BLIMP-1. BCL-6 is a repressor of *MYC* in the germinal center B cells, whereas BLIMP-1 is a repressor of *MYC* in

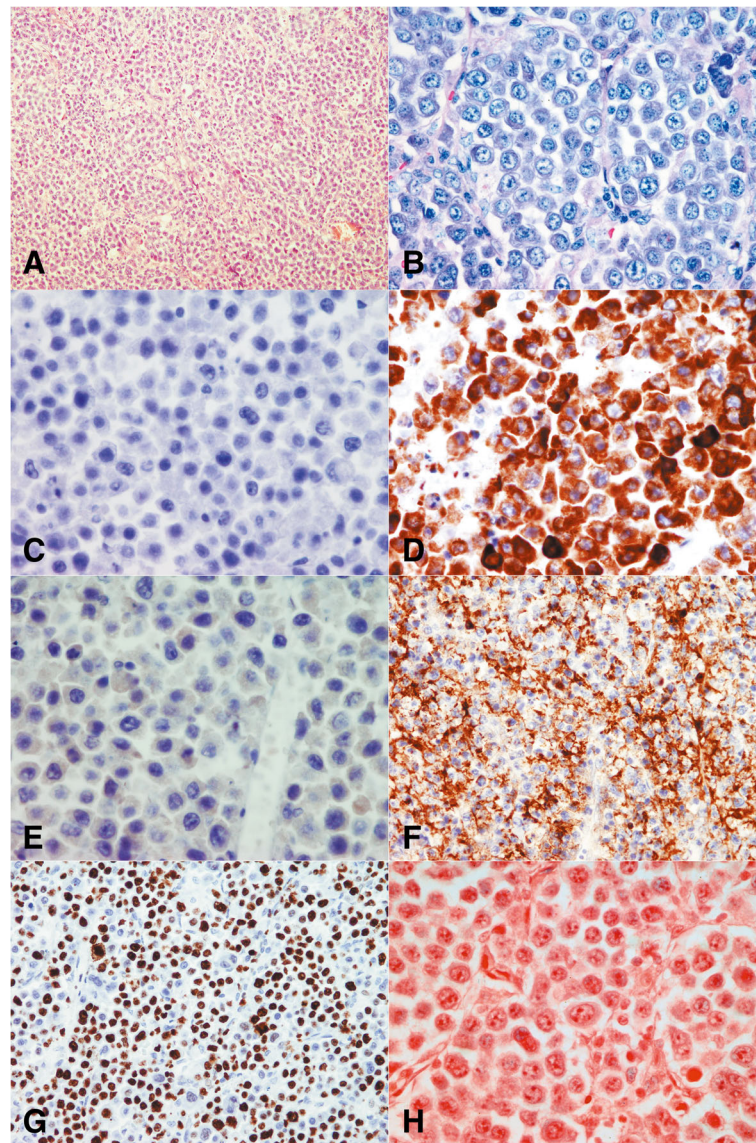


Fig. 2 Histology and immunohistochemistry of plasmablastic lymphoma. **a** Haematoxylin and eosin section shows monotonous, lymphoid cells with “starry sky” pattern (magnification $\times 100$). **b** Giemsa staining highlights the plasmablastic appearance with eccentrically positioned nuclei, prominent nucleoli and abundant basophilic cytoplasm (magnification $\times 400$). **c-f** Atypical lymphoid cells were negative for CD20 (**c**) and CD30 (**e**), while they were positive for CD79a (**d**) and CD138 (**f**) (immunoperoxidase stain; **c, d** and **e** magnification $\times 400$; **f** magnification $\times 200$). **g** Nuclear proliferation rate, assessed by Ki-67 was approximately 80% (immunoperoxidase stain; magnification $\times 200$). **h** EBV encoded RNA (EBER) was negative in the nuclei of the atypical cells (magnification $\times 400$)

terminally differentiated B cells [2]. According to one theory, *MYC* translocation may lead to the plasmablastic morphology and create a more aggressive disease state [3, 19]. Also, it has been observed that *MYC* rearrangements are more often seen in EBV-positive compared to EBV-negative tumors [18]. Therefore, we also performed c-*MYC* immunohistochemistry which showed strong protein expression in >40% of tumor cells (Fig. 3). Furthermore, to examine c-*MYC* gene status, fluorescent in situ hybridization (FISH) was performed using Vysis *MYC* Break Apart FISH Probe Kit (Abbott Molecular, IL,

USA) and Vysis LSI *IGH/MYC*, CEP 8 Tri-color Dual Fusion Translocation Probe (Abbott Molecular, IL, USA). We found 8q24 rearrangement within tumor cells but we did not find specified t (8,14) *IGH/MYC* translocation (Fig. 3).

The present case was negative for HIV and EBV infection was not detected by ISH. Some studies revealed that EBV infection was detected in only 17% of HIV-negative PBL cases, suggesting that EBV is not a distinct contributor in the pathogenesis of PBL in patients without HIV infection [20, 21].

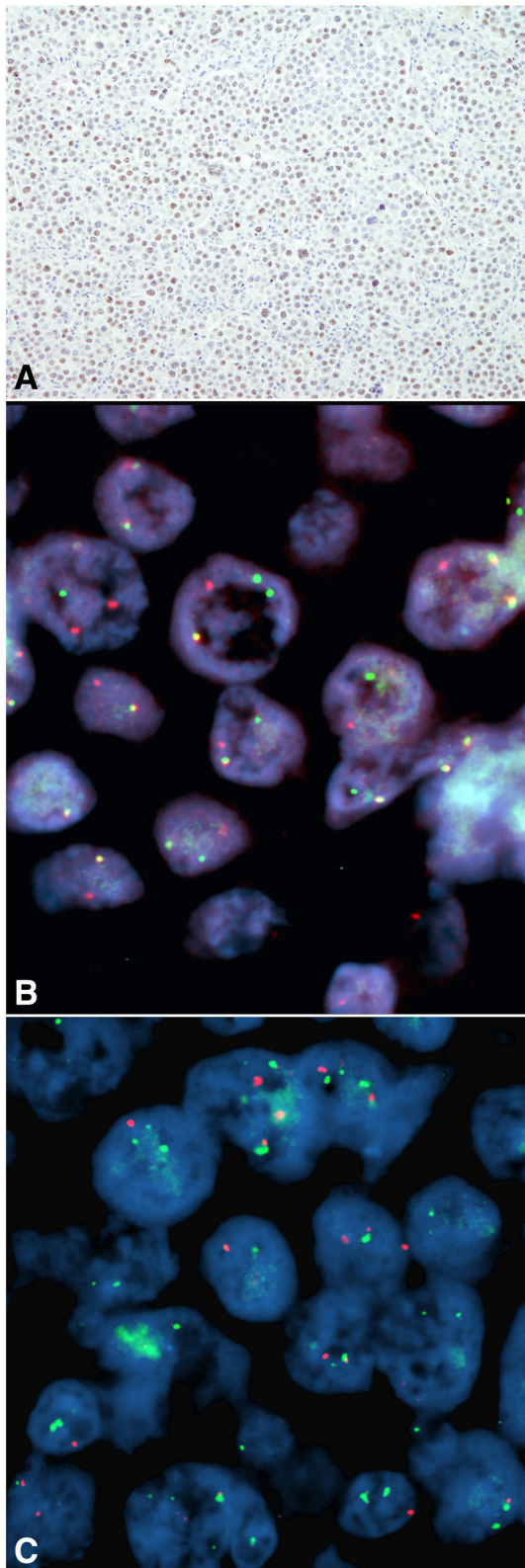


Fig. 3 MYC immunohistochemistry and fluorescent in situ hybridization (FISH). **a** Large atypical plasmablasts were c-MYC positive in >40% of cells. **b** FISH analysis using Vysis MYC Break Apart FISH Probe Kit revealed 8q24 rearrangement within tumour cells (cells without rearrangement should have 2 yellow fusion signals; magnification $\times 1000$). **c** FISH analysis using Vysis LSI IGH/MYC, CEP 8 Tri-color Dual Fusion Translocation Probe is negative for t(8,14), without yellow signals specific for IGH/MYC translocation (normal 2 green (IGH) and 2 red (MYC, 8q24) signals); magnification $\times 1000$

The minimum morphologic criteria, required to diagnose PBL, are monomorphic cellular proliferation of plasmablasts, with either centrally or eccentrically placed nuclei, with high nuclear-cytoplasmic rate, a moderate amount of eosinophilic cytoplasm, a high mitotic index and the absence of plasma cells in the background [22, 23]. High mitotic activity found in all PBLs as determined by Ki-67 proliferation index, is consistent with our case. PBL should also be differentiated from other malignant diseases including DLBCL, poorly differentiated carcinomas, melanoma, gastrointestinal stromal tumors, Burkitt's lymphoma and anaplastic large cell lymphoma. However, the main differential diagnosis of PBL is plasmablastic or anaplastic multiple myeloma. In practice the distinction between these frequently depends on clinical correlation. HIV infection, immune suppression and EBER positivity favor PBL diagnosis, while the presence of monoclonal paraproteinemia, lytic bone lesions, hypercalcemia and renal dysfunction favors a diagnosis of myeloma. Also high Ki-67 proliferation index and CD56 positivity favors diagnosis of PBL over myeloma [6].

PBL is a highly aggressive disease with the majority of patients dying within the first year after initial presentation. A systemic review of HIV-positive patients with PBL showed a median overall survival (OS) of 15 months versus 9 months in HIV-negative patients [24]. Furthermore, very high Ki-67 index and MYC gene rearrangements, have been shown to be associated with shorter OS in patients with PBL [2, 21]. Also extra-oral PBL are more frequently disseminated at the time of diagnosis (57% of patients are at stage IV) [21]. The prognostic value of EBV related antigens expression in PBL cases is unclear. Some studies have shown that EBV is not associated with outcome in HIV-associated PBL while other studies have shown that EBV infection correlated with better outcome in immune competent patients [2]. In our case, the patient's immune competent, HIV-negative status, EBER negativity, extra-oral location, high Ki-67 proliferation index and MYC rearrangement were all predictors of the aggressive clinical course and short survival.

PBL remains difficult to diagnose and to treat. A standard therapeutic approach for patients with PBL has not been established. In particular, the use of cyclophosphamide, doxorubicin, vincristine and prednisone (CHOP) is considered inadequate therapy, and current

guidelines recommend more intensive treatments [2]. Such treatments include infusional etoposide, vincristine and doxorubicin with bolus cyclophosphamide and prednisone (EPOCH), cyclophosphamide, vincristine, doxorubicin, methotrexate alternating with ifosfamide, etoposide and cytarabine (CODOX-M/IVAC) [25], or hyperfractionated cyclophosphamide, vincristine, doxorubicin, and dexamethasone alternating with methotrexate and cytarabine (hyper-CVAD) [26]. More recently, the role of stem cell transplantation (SCT) in patients with PBL has been assessed [27]. It appears patients with PBL with chemotherapy-sensitive disease might benefit from autologous SCT in the first remission therefore, it seems rational to explore the use of autologous SCT earlier in the disease course [27]. Some studies have reported that bortezomib (proteasome inhibitor) alone or in combination with chemotherapy may have an antitumor effect in PBL considering it has many morphologic and immunophenotypic similarities with myeloma [28]. In our case, the patient received 4 cycles of DA-EPOCH, followed by 1 cycle of salvage DHAP and 4 cycles of alternating modified CODOX-M/IVAC chemotherapy. Unfortunately autologous SCT could not be performed due to the patient's worsening clinical status.

Conclusions

In conclusion, we report a peculiar, extra-oral PBL case detected within a mature cystic teratoma in an HIV-negative, immune competent patient. To the best of our knowledge, this is the first case of PBL described in a mature ovarian cystic teratoma. Diagnosis of PBL is challenging, especially when it arises in younger, HIV-negative, immune competent, female patients without EBER positivity. Currently there is no definitive treatment regimen capable of providing curative results, therefore until standardized chemotherapy is identified, management of patients with PBL should be on case to case basis [29].

Additional file

Additional file 1: Timeline table Hadžisejdić et al. case report. (DOC 39 kb)

Abbreviations

AST: Aspartate aminotransferase; CD: Cluster of differentiation; CRP: C-reactive protein; DLCL: Diffuse large B-cell lymphoma; EBER: EBV-encoded RNA; EBV: Epstein-Barr virus; FISH: fluorescent in situ hybridization; HBV: Hepatitis B virus; HCV: Hepatitis C virus; HIV: Human immunodeficiency virus; ISH: In situ hybridisation; LDH: Lactate dehydrogenase; MSCT: Multi-slice computed tomography; NHL: Non-Hodgkin lymphoma; OS: Overall survival; PBL: Plasmablastic lymphoma; PET-CT: Positron emission tomography and computed tomography; SCT: Stem cell transplantation; US: Ultrasound

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Availability of data and materials

The datasets used and/or analyzed during the current study available from the corresponding author on the reasonable request.

Authors' contributions

IH major contributor in writing the manuscript, collecting clinical data, histology and immunohistochemistry analyses, performed FISH analyses; EB clinical data collection and performed histological examination; LV clinical data collection, figure and manuscript formatting; ADN therapy related data collection and writing; KL performed histological examination, manuscript preparation suggestions; MK performed histological examination, manuscript preparation suggestions; NJ immunohistochemistry interpretation and FISH analyses, final manuscript revision. All authors read and approved the final manuscript.

Ethics approval and consent to participate

All the clinical data obtained and tissue analyses were handled with the care and respect to the patient's anonymity and approved by Ethical Committees of the Rijeka University Hospital Centre and Faculty of Medicine, University of Rijeka.

Consent for publication

Written informed consent was obtained from the patient's legal guardian for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Competing interests

The authors declare that they have no competing interests.

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References

- McKenna RW, Kyle RA, Kuehl WM, Grogan TM, Harris NL, Coupland RW. Plasmablastic lymphoma. In: Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, et al., editors. WHO classification of Tumors of hematopoietic and lymphoid tissues. 4th ed. Lyon: IARC; 2008. p. 256–7.
- Castillo JJ, Bibas M, Miranda RN. The biology and treatment of plasmablastic lymphoma. *Blood*. 2015;125(15):2323–30.
- Castillo JJ, Reagan JL. Plasmablastic lymphoma: a systematic review. *Sci World J*. 2011;11:687–96.
- Wilson WH, Grossbard ML, Pittaluga S, Cole D, Pearson D, Drbohlav N, Steinberg SM, Little RF, Janik J, Gutierrez M, Raffeld M, Staudt L, Cheson BD, Longo DL, Harris N, Jaffe ES, Chabner BA, Wittes R, Balis F. Dose-adjusted EPOCH chemotherapy for untreated large B-cell lymphomas: a pharmacodynamic approach with high efficacy. *Blood*. 2002;99(8):2685–93.
- Delecluse HJ, Anagnostopoulos I, Dallenbach F, Hummel M, Marafioti T, Schneider U, Huhn D, Schmidt-Westhausen A, Reichart PA, Gross U, Stein H. Plasmablastic lymphomas of the oral cavity: a new entity associated with the human immunodeficiency virus infection. *Blood*. 1997;89(4):1413–20.
- Cao C, Liu T, Lou S, Liu W, Shen K, Xiang B. Unusual presentation of duodenal plasmablastic lymphoma in an immunocompetent patient: a case report and literature review. *Oncol Lett*. 2014;8(6):2539–42.
- Cha JM, Lee JJ, Joo KR, Jung SW, Shin HP, Lee JJ, Kim GY. A case report with plasmablastic lymphoma of the jejunum. *J Korean Med Sci*. 2010;25(3):496–500.
- Vang R, Medeiros LJ, Warnke RA, Higgins JP. Ovarian non-Hodgkin's lymphoma: a clinic pathologic study of eight primary cases. *Mod Pathol*. 2001;14(11):1093–9.
- Maguire A, Castriciano G, Walker J, Molloy K, Quinn F, Osman N, Vandenberghe E, Jeffers M. Case study: diffuse large B-cell lymphoma arising in ovarian mature cystic teratoma. *Int J Gynecol Pathol*. 2015;34(5):459–64.
- Valli R, Froio E, Alvarez de Celis MI, Mandato VD, Piana S. Diffuse large B-cell lymphoma occurring in an ovarian cystic teratoma: expanding the spectrum

- of large B-cell lymphoma associated with chronic inflammation. *Hum Pathol.* 2014;45(12):2507–11.
11. Tandon N, Sultana S, Sun H, Zhang S. Follicular lymphoma arising in a mature cystic teratoma in a 26 year old female. *Ann Clin Lab Sci.* 2016;46(3):298–301.
 12. Dimopoulos MA, Daliani D, Pugh W, Gershenson D, Cabanillas F, Sarris AH. Primary ovarian non-Hodgkin's lymphoma: outcome after treatment with combination chemotherapy. *Gynecol Oncol.* 1997;64:446–50.
 13. Monterroso V, Jaffe ES, Merino MJ, Medeiros LJ. Malignant lymphoma involving the ovary. A clinicopathologic analysis of 39 cases. *Am J Surg Pathol.* 1993;17:154–70.
 14. Vortmeyer AO, Devouassoux-Shisheboran M, Li G, Mohr V, Tavassoli F, Zhuang Z. Microdissection-based analysis of mature ovarian teratoma. *Am J Pathol.* 1999;154:987–91.
 15. Fox H, Langley FA, Govan AD, Hill AS, Bennett MH. Malignant lymphoma presenting as an ovarian tumor: a clinicopathological analysis of 34 cases. *Br J Obstet Gynecol.* 1988;95:386–90.
 16. Dawson MA, Schwarzer AP, McLean C, Oei P, Campbell LJ, Wright E, Shortt J, Street AM. AIDS-related plasmablastic lymphoma of the oral cavity associated with an IGH/MYC translocation—treatment with autologous stem-cell transplantation in a patient with severe haemophilia-a. *Haematologica.* 2007;92(1):e11–2.
 17. Bogusz AM, Seegmiller AC, Garcia R, Shang P, Ashfaq R, Chen W. Plasmablastic lymphomas with MYC/IgH rearrangement: report of three cases and review of the literature. *Am J Clin Pathol.* 2009;132(4):597–605.
 18. Valera A, Balagué O, Colomo L, Martínez A, Delabie J, Taddesse-Heath L, Jaffe ES, Campo E. IG/MYC rearrangements are the main cytogenetical teration in plasmablastic lymphomas. *Am J Surg Pathol.* 2010;34(11):1686–94.
 19. Taddesse-Heath L, Meloni-Ehrig A, Scheerle J, Kelly JC, Jaffe ES. Plasmablastic lymphoma with MYC translocation: evidence for a common pathway in the generation of plasmablastic features. *Mod Pathol.* 2010;23(7):991–9.
 20. Lorsbach RB, Hsi ED, Dogan A, Fend F. Plasma cell myeloma and related neoplasms. *Am J Clin Pathol.* 2011;136(2):168–82.
 21. Wang HW, Yang W, Sun JZ, Lu JY, Li M, Sun L. Plasmablastic lymphoma of the small intestine: case report and literature review. *World J Gastroenterol.* 2012;18(45):6677–81.
 22. Bibas M, Castillo JJ. Current knowledge on HIV-associated Plasmablastic lymphoma. *Mediterr J Hematol Infect Dis.* 2014;6(1):e2014–64.
 23. Kane S, Khurana A, Parulkar G, Shet T, Prabhaskar K, Nair R, Gujral S. Minimum diagnostic criteria for plasmablastic lymphoma of oral/sinonasal region encountered in a tertiary cancer hospital of a developing country. *J Oral Pathol Med.* 2009;38(1):138–44.
 24. Castillo JJ, Winer ES, Stachurski D, Perez K, Jabbour M, Milani C, Colvin GA, Butera JN. HIV-negative plasmablastic lymphoma: not in the mouth. *Clin Lymphoma Myeloma Leuk.* 2011;11(2):185–9.
 25. Mead GM, Sydes MR, Walewski J, Grigg A, Hatton CS, Pescosta N, Guarnaccia C, Lewis MS, McKendrick J, Stenning SP, Wright D, UKLG LY06 collaborators. An international evaluation of CODOX-M and CODOX-M alternating with IVAC in adult Burkitt's lymphoma: results of United Kingdom lymphoma group LY06 study. *Ann Oncol.* 2002;13(8):1264–74.
 26. Cortes J, Thomas D, Rios A, Koller C, O'Brien S, Jeha S, Faderl S, Kantarjian H. Hyperfractionated cyclophosphamide, vincristine, doxorubicin, and dexamethasone and highly active antiretroviral therapy for patients with acquired immunodeficiency syndrome-related Burkitt lymphoma/leukemia. *Cancer.* 2002;94(5):1492–9.
 27. Al-Malki MM, Castillo JJ, Sloan JM, Re A. Hematopoietic cell transplantation for plasmablastic lymphoma: a review. *Biol Blood Marrow Transplant.* 2014;20(12):1877–84.
 28. Saba NS, Dang D, Saba J, Cao C, Janbain M, Maalouf B, Safah H. Bortezomib in plasmablastic lymphoma: a case report and review of the literature. *Onkologie.* 2013;36(5):287–91.
 29. Saraceni C, Agostino N, Cornfield DB, Gupta R. Plasmablastic lymphoma of the maxillary sinus in an HIV-negative patient: a case report and literature review. *Spring.* 2013;2(1):142.

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