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Source / Izvornik: Marine Pollution Bulletin, 2018, 131, 95 - 105

Journal article, Accepted version Rad u časopisu, Završna verzija rukopisa prihvaćena za objavljivanje (postprint)

https://doi.org/10.1016/j.marpolbul.2018.03.002

Permanent link / Trajna poveznica: https://urn.nsk.hr/um:nbn:hr:184:650766

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A baseline study of the metallothioneins content in digestive gland of the Norway lobster
 Nephrops norvegicus from Northern Adriatic Sea: Body size, season, gender and metal specific
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Keywords: *Nephrops norvegicus*, metallothioneins, metals, reactive oxygen species,
 energy reserves, biomarker baseline

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ABSTRACT

Metallothioneins content was investigated in digestive gland of two wild-caught Norway 25 lobster Nephrops norvegicus populations from the Northern Adriatic Sea, in relation to body 26 size, season and gender. Concomitant accumulation of cadmium, mercury, arsenic, lead, 27 28 chromium and manganese, reactive oxygen species concentration and energy reserves in 29 digestive gland were also assessed. While differences between genders were not recorded, metallothioneins content seasonal trends were affected by body size. Most of parameters 30 displayed inconsistent trends across sampling sites. Significant correlation between 31 metallothioneins content and cadmium, arsenic and mercury concentrations was recorded only 32 33 for larger lobsters. A negative correlation of reactive oxygen species concentration and metallothioneins content was observed for small, but not large lobsters. Energy reserves, in 34 particular lipids, could considerably influence biochemical and chemical parameters variations. 35 The present results constitute the essential baseline for future studies aimed at evaluating the N. 36 norvegicus health in relation to metal contamination of coastal sediments. 37

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The Norway lobster *Nephrops norvegicus* (Linneaus, 1758) is among the most economically important crustacean species of the Mediterranean Sea and NE Atlantic. For the Adriatic Sea in particular, the overall population of *N. norvegicus* has been dramatically declining due to excessive exploitation of fishing grounds (Piccinetti et al., 2012). Besides, coastal zones may be subjected to intensive anthropogenic activities, such as increasing urbanization and industrialization. The resulting contamination with metals in particular represents a significant risk for aquatic biota, given their ubiquity, long-term persistence in the sediment and potential toxicity to benthic organisms. However, the assessment of contaminants based on chemical analyses in seawater and sediments is still predominant and little information is available regarding toxic effect and detoxification of metals for burrowing lifestyle crustaceans such as *N. norvegicus*.

50 Metallothioneins (MTs) represent a group of low molecular weight, cysteine-rich proteins 51 essential for metal detoxification and homeostasis due to their ability of binding and sequestering several metals. Previous studies showed MTs responsiveness to elevated metal 52 exposure in both invertebrates and vertebrates and prompted their application as biomarker of 53 early metal stress in many marine organisms (Amiard et al., 2006). Even though the utility of 54 MTs as biomarker of metal exposure was found to be uncertain or doubtful for some crustaceans 55 (Legras et al., 2000; Ortega et al., 2017; Pedersen et al., 2014), evidence of increased expression 56 and synthesis of MTs upon in vivo laboratory exposure to metals, primarily to cadmium, were 57 reported for decapode crustaceans Panularis argus, Charybdis japonica and N. norvegicus 58 59 (Canli et al., 1997; Moltó et al., 2005; Pan and Zhang, 2006). Essential metals such as Cu and Zn are also effective MTs inducers in crustaceans (Barka et al., 2001; Legras et al., 2000). Less 60 information is available on MTs induction by other metals, but the increase of MTs content in 61 crustaceans was recorded after laboratory exposure to mercury (Barka et al., 2001; 2007) and 62 arsenic (Vellinger et al., 2013). Field surveys also revealed positive correlations of MTs content 63 64 with As and Hg as well as with Pb, Cr and Mn (Faria et al., 2010; Lavadras et al., 2014; Martin-Díaz et al., 2009). 65

In addition to the metal-detoxifying role, crustaceans MTs are also involved in protection from reactive oxygen species (ROS) that can cause damage to cell macromolecules when produced in excessive quantities (Pan and Zhang, 2006; Moltó et al., 2007; Lobato et al., 2013; Felix-Portillo et al., 2014). Metals can be distributed into soluble cytosol, or prevail within insoluble fraction that include metal-rich granules, involved in the second important metal detoxification pathway in crustaceans (Barka, 2007; Legras et al., 2000; Mouneyrac et al., 2001; Nunez-Nogueira et al., 2010).

73 While controlled laboratory-based exposure studies provided substantial evidence on the direct link of toxic chemicals and biomarkers response, field studies of anthropogenic 74 contamination effects still represent a considerable challenge for the scientific community due 75 to limited knowledge on natural variability of biomarkers in sentinel marine organisms. 76 77 Confounding factors such as intrinsic biotic and environmental parameters cause large amplitudes of biomarkers' response that could consequently be overestimated and incorrectly 78 attributed to the toxic effect of contaminants. Thus, definition of natural biomarker variation 79 range using data collected from reference sites has become a priority task of ecotoxicological 80 studies as a foundation for realistic evaluation of contaminants effect in the field (Barrick et al., 81 2016; Davies and Vethaak, 2012 and references therein). Fluctuations of MTs content related 82 to gender, season and reproductive status were already identified in the tissues of some 83 decapode crustacean species (Chiodi Boudet et al., 2013; Giarratano et al., 2016; Lavadras et 84 85 al., 2014; Maria et al., 2009; Mouneyrac et al., 2001). Consequently, in order to avoid possible misinterpretation of contaminants' effect in the natural habitat it is essential to establish the 86 MTs baseline levels in relation to potentially confounding factors. 87

The aim of this study was to investigate the MTs content in digestive gland of N. 88 norvegicus with respect to body size, season, gender and metals concentrations. In addition, 89 ROS level and energy reserves were assessed as potential source of MTs content variability. 90 Samples were obtained from two fishing areas of the Kvarner bay (Northern Adriatic Sea, 91 Croatia). Since in general, the investigations of anthropogenic impact have been mostly 92 concentrated along the coastal line, data concerning biological effect of potential contaminants 93 94 in species from off-shore marine habitats is currently scarce and fragmented. The results of the present study will represent the necessary prerequisite for future studies oriented towards 95 evaluation of the risk of metal contamination for benthic crustaceans. 96

97 Kvarner Bay is a semi - enclosed and relatively shallow coastal area on the North eastern 98 part of Adriatic Sea (Croatia). Sampling site S1 wass located in the inner part of the bay, closer to the Rijeka harbour (~8 - 11 km) that is the major source of metals contamination. The second 99 sampling site S2 was positioned approximately 70 km southern from Rijeka harbour and is 100 more influenced by open sea circulation (Fig. 1). Data on the level of metals in the sediment 101 102 and biota for sampling sites is currently unavailable. Although generally improving trends have been recently detected for coastal zones of the Kvarner Bay, elevated levels of metals were 103 occasionally recorded in the sediments of various near shore locations, mostly within areas 104 adjacent to urbanized areas and zones of intensive industrial activity, whereas sediments from 105 sites furthest from the coast displayed substantially lower values (Cukrov et al., 2011, 2014). 106 107 Sediment metal enrichment of various offshore areas of the Kvarner Bay corresponds to that of unpolluted sites within central and southern Adriatic far from any known anthropogenic sources 108 (Ilijanić et al., 2014). Nevertheless, the possibility of accidental discharge cannot be ruled out 109 since both sampling sites are intersected with important transport routes for cargo and touristic 110 111 ships. In addition, the northern Adriatic is enriched with mercury predominantly originating from inland deposits and driven by sea currents towards the rest of the Adriatic Sea (Kotnik et 112 113 al., 2015).

Specimens of Nephrops norvegicus were collected in autumn 2014 and spring 2015 by 114 bottom trawl fishing gear. Trawling depths for sites S1 and S2 were at 55-62 and 70-78 m, 115 respectively. Immediately following capture healthy and undamaged animals were selected and 116 transported to the laboratory in thermally isolated containers. The gender of organisms was 117 determined by checking the morphology of the first pair of abdominal swimmeret (pleopodes) 118 that are thicker and rigid in males. The carapace length (CL; from eye socked to mid hind edge 119 of carapace) was recorded for each lobster. For each site, season and gender the organisms were 120 subsequently classified into two non-overlapping and relatively well-defined body-size group 121 according to their CL (Table 1). Lobsters of CL below 36 mm were considered as "small" 122 whereas the second body-size group, hereafter referred to as "large", comprised all lobsters 123 samples of CL larger than 36 mm due to heterogeneity of CL values and difficulties in obtaining 124 a balanced gender ratio. The total of 144 specimens of N. norvegicus were dissected to obtain 125 digestive gland tissue samples that were immediately frozen in liquid nitrogen and stored at -126 80°C. 127

Portions of 0.5 g of freeze dried digestive gland tissue samples were digested using Anton
Paar Multiwave 3000 microwave system (Perkin Elmer, USA) equipped with pressurized
vessels, using 5 mL of 65% nitric acid per sample (HNO₃ Suprapur, Merck, Germany), over a
20 minutes operation cycle at 200 °C. Digested samples were transferred to 25 ml volumetric

flasks and added with ultrapure water (Siemens). The concentrations of Cd, As, Pb, Cr and Mn 132 were determined using the inductively coupled plasma mass spectrometer, ICP MS NexION 133 300X, equipped with S10 autosampler (Perkin Elmer, USA). Multielement solution (NexION 134 Setup Solution, Perkin Elmer, USA) was used as tuning solution, covering a wide range of 135 masses of elements. Multielement standard solution (Perkin Elmer, USA) was used to prepare 136 calibration curves. The calibration curves with $R^2 > 0,999$ were accepted for concentration 137 138 calculation. For each experiment, a run included blank, certified reference material (CRM) and samples which were analysed in triplicate to eliminate any batch-specific error. The method for 139 measurement of Cd, As, Pb, Cr and Mn was validated using IAEA-407 reference material (fish 140 tissue) (International Atomic Energy Agency, Austria). The mean recovery values for Cd, As, 141 Pb, Cr and Mn were 97%, 89%, 105%, 110% and 108%, respectively. For Hg determination, 142 approx. 0.1 g of freeze dried digestive gland tissue was weighed, transferred directly into 143 analyser's vessel and analysed using the atomic absorption spectrometer AMA 254 (Advanced 144 Mercury Analyser, Leco, USA). Single element standard solution for Hg was used for 145 146 instrument calibration (LGC Standards, USA). The method for Hg measurement was validated using NIST 2976 reference material (mussels tissue) (National Institute of Standards and 147 Technology, USA). The mean recovery for Hg was 104%. The concentrations of all metals 148 were expressed as µg per g of tissue dry weight (d.w.). Data for Cu and Zn concentrations were 149 150 provided by Glad (personal communication).

Measurement of MTs content in digestive gland was performed in accordance to the method of Viarengo et al. (1997). MTs content was determined in a partially purified low molecular weight metalloproteins fraction following acidic ethanol/chloroform extraction of the homogenate, by spectrophotometric measurement at 412 nm. For the calculation of MTs content, serial dilutions of reduced glutathione (GSH) were used as reference standard, assuming the content of 18 Cys residues (Zhu et al., 1994). MTs concentration was expressed as nmoles of GSH per g of wet tissue weight (w.w.).

The OxiSelectTM In Vitro ROS/RNS Assay Kit (Cell Biolabs, inc. USA) was used for the 158 159 measurement of total free radicals in the samples, according to the manufacturer instructions. The method is based on the measurement of 2', 7'- dichlorodihydrofluorescein (DCF) 160 fluorescence that results from oxidation of highly reactive dichlorodihydrofluorescein (DCFH). 161 The intensity of DCF fluorescence that is proportional to total free radicals level in the sample 162 was measured using the fluorescence plate reader Fluoroscan AscentTM (Labsystem, Finland) 163 at 480 nm excitation/530 nm emission. For calculation of the free radical content, serial 164 dilutions of H₂O₂ were used, and the concentration of ROS/RNS was expressed as mmol/mg of 165 tissue wet weight (w.w.). 166

Quantitative determination of total lipids in digestive gland tissue was performed by sulfo-167 phospho-vanillin colorimetric method (Glad, 2017). Briefly, lipids were extracted by vigorous 168 mixing of freeze dried digestive gland tissue portions (10-20 mg) with chloroform/methanol 169 2:1 (V/V) solution, followed by incubation at +4°C for 10 min and centrifugation at 4000 rpm 170 for 10 min. Aliquots of each sample were evaporated to dryness under nitrogen stream in glass 171 172 tubes, vigorously mixed with concentrated sulphuric acid and heated at 100°C for 10 minutes. The total lipid content was determined in cooled samples using phospho-vanillin reagent and 173 serial dilutions of cholesterol standards. The absorbance values of blank and samples were 174 recorded at 490 nm and the concentration was expressed as % of lipids per g of tissue dry weight 175

(d.w.). For determination of proteins in digestive gland tissue, portions of freeze dried tissue
(10-20 mg) were mixed with 10% sodium hydroxide and incubated for 1 hour at 100°C to digest
proteins. Samples were then subjected to Bradford assay (Bradford, 1976) with bovine serum
albumin as protein standard. The absorbance values of blank and samples were recorded at 595
nm. Concentration was expressed as % of proteins per g of tissue dry weight (d.w.).

Data are graphically presented as box and whisker plots (small, N=6; large, N=12). Since 181 182 the Shapiro-Wilk and Levene's tests revealed that requirements of normality and homogeneity of variance were not met, the non-parametric Kruskal Wallis and Mann-Whitney's U-test were 183 used for statistical analyses. The relationships between MTs content and metal concentrations, 184 ROS and energy reserves level were determined using Spearman's rank correlation analysis. 185 186 Differences were considered significant at p < 0.05. Data for MTs content of all samples was used for frequency distribution histogram. Principal component analysis (PCA) was performed 187 in order to investigate the overall data pattern. The background level was calculated as the mean 188 value of MTs content of all 144 samples in total. All the analyses were performed using 189 190 RStudio, version 0.98.1028 (RStudio Team, 2015).

For each body size group of *N. norvegicus*, data on season and gender specific MTs content, metals and ROS concentrations and energy reserves level in digestive gland tissue were presented separately for sites S1 and S2.

194 Content of MTs in digestive gland of small and large Nephrops norvegicus ranged from 1.1 to 44.4 and from 7.2 to 39.8 nmol/g w.w., for sites S1 and S2, respectively (Fig. 2). Values 195 for MTs content varied significantly between seasons at site S1, with a contrasting trend 196 displayed between body size groups. At site S2, MTs content was significantly higher in spring 197 only for small male lobsters. Differences between small and large lobsters were also found at 198 199 site S1 (Table 2). Significant differences in MTs content were observed between sites, being higher at site S1 in autumn for small males of both genders and in spring for large females. The 200 opposite pattern was observed at site S2 (Table 3). Concentration of Cd ranged from 3 to 28.1 201 $\mu g/g$ d.w. and from 4.1 to 37.9 $\mu g/g$ d.w. for lobsters from sites S1 and S2, respectively (Fig. 202 203 3). At site S1, Cd values displayed marked seasonality for both body size classes, being significantly higher predominantly in spring. Gender dependent differences were not recorded. 204 At site S2, no seasonality could be observed. Concentrations of Cd did not show differences 205 with respect to body size (Table 2). Values recorded at S2 were occasionally significantly 206 207 higher than at site S1, predominantly in autumn (Table 3). Concentration of As ranged from 33.6 to 594.4 µg/g d.w. and from 61.1 to 1254.1 µg/g d.w., at sites S1 and S2, respectively (Fig. 208 3). Gender and season dependent As accumulation was detected for both body size groups and 209 sites, mainly being significantly higher in spring and in males. Significantly higher As levels 210 were found in smaller males, except in spring at site S2 (Table 2). Concentration of As was 211 almost regularly significantly higher at site S2 (Table 3). Concentration of Hg ranged from 1 to 212 5.3 μ g/g d.w. and from 0.9 to 5.6 μ g/g d.w., at sites S1 and S2, respectively (Fig. 3). 213 Significantly higher Hg concentration was recorded almost regularly in spring, predominantly 214 in males. Differences related to body size, although sometimes significant, did not express a 215 clear trend (Table 2). Concentrations of Hg were often significantly higher at site S1, 216 particularly in spring (Table 3). Concentrations of Pb ranged from 0.04 to 3.6 µg/g d.w. and 217 from 0.06 to 1.6 µg/g d.w. at sites S1 and S2, respectively (Fig. 3). At site S1, Pb levels 218 displayed similar trends, but significantly higher concentrations were recorded predominantly 219

in male lobsters and in spring. Opposite seasonal patterns with higher autumn values were 220 observed at site S2 for both body size categories of females only. Concentration of Pb was 221 significantly higher in small lobsters of both genders at site S1 in spring, and at site S2 in 222 females in autumn (Table 2). Differences of Pb accumulation between sites were season-223 specific, that is, significantly higher at S1 and S2 in spring and autumn, respectively (Table 3). 224 Concentration of Cr ranged from 1.1 to 8 μ g/g d.w. and from 1.1 to 5.45 μ g/g d.w. at sites S1 225 226 and S2, respectively (Fig. 3). The season related pattern was more consistent at site S2, where significantly higher Cr concentrations were recorded in autumn. There were no differences 227 between genders. Significant differences were detected between body size groups only for 228 males at site S1, displaying higher Cr concentrations in large and small lobsters in autumn and 229 230 spring, respectively (Table 2). Concentration of Cr was significantly higher at S1 only in spring (Table 3). The range of Mn concentrations for lobsters from sites S1 and S2 was between 7 to 231 33.7 µg/g d.w. and from 7.6 to 42.2 µg/g d.w., respectively (Fig. 3). Seasonal differences were 232 recorded only for females of both body size groups, displaying significantly higher levels in 233 234 spring at site S1, and in autumn at site S2 (Table 2). Values for Mg concentration were significantly higher in small lobsters, but almost exclusively at site S1 (Table 2). Accumulation 235 of Mn was significantly higher at site S2 for females only (Table 3). 236

Values for ROS varied between 0.8 and 16.9 mmol/ mg w.w. and 1.4 and 12.4 mmol/ mg
w.w. at sites S1 and S2, respectively (Fig. 4). The pattern of ROS level was inconsistent, but
gender and season dependent significant differences were observed at both sites. Significantly
higher concentration of ROS in small lobsters was occasionally found (Table 2). Values for
ROS were regularly significantly different between sites, being higher predominantly at S2,
while the opposite trend was detected only for males in spring (Table 3).

The values for lipid content varied between 20 and 50 % d.w. and between 24 and 37 % d.w., at sites S1 and S2, respectively. Lipid content was elevated in autumn (Table 4). A gender dependent lipid content could be discerned only at site S1, being higher in female lobsters from both body size groups. Total protein content at sites S1 and S2 ranged from 18 to 31 % d.w. and 18 and 35 % d.w., respectively. No consistent pattern could be detected at either of the two sites.

A significant negative correlation was detected between MTs and metals (r_s= Mn, -0.4; Cd, 249 -0.46, Cu, -0.46; Zn, -0.41, p<0.05) for small lobsters (Table 5). Large organisms displayed 250 significant (p<0.05) positive correlation with Cd ($r_s=0.25$), Hg ($r_s=0.31$) and As ($r_s=0.37$). 251 Content of MTs was negatively correlated to ROS (r_s =-0.45, p<0.05) in digestive gland of small 252 lobsters only. Large, but not small lobsters, displayed significant negative correlation with 253 lipids (r_s =-0.31, p<0.05). Significant correlations (p<0.05) were recorded between metal 254 concentrations and lipids, namely for Hg, Pb, Mn and Cr (r_s =-0.38 to -0.67) and for Cd, As, Pb 255 Hg, Mn and Cu (r_s = -0.3 to -0.56) in digestive gland of small and large lobsters, respectively. 256

A principal component analysis (PCA) was performed using data on MTs content, ROS level, metals accumulation and energy reserves, obtained from all individuals sampled of both body size groups, seasons, genders and sites. The first two principal components PC1 and PC2 accounted for 59.2% and 51.6% for small and large lobsters, respectively (Fig. 5, upper and lower panel). Generally large variation was observed for samples spread on both PCA ordination plots. The contribution of the variables in each of the first two PCs varied between two body size groups. With exception of site S1 in autumn, small lobsters were mostly grouped

to the left part of the PC1, with contribution of Cd, As, Cu, Zn concentrations and ROS level 264 that were negatively correlated to PC1, while MTs content was positively correlated to PC1. 265 Small males and females from site S1 in autumn were separated along PC2 as a result of the 266 differences in the accumulation of Mn, Cr, Pb and lipids content (negatively correlated to PC2). 267 Large males and females from site S1 sampled in autumn were distributed at the left side of 268 269 PC1 and had generally lower levels of metals and MTs content and higher lipids content. Large 270 lobsters from site S2 were roughly separated along PC2, with spring samples being located in its positive side due to high accumulation of Hg and As, and autumn sample in the negative 271 side due to association with higher ROS level and Cr and Zn concentrations. 272

Figure 6. represents the frequency distribution histogram for MTs content of all 144 samples in total. The preliminary threshold value was defined as mean $+ 1\sigma$ and expressed the value of 25.2 nmol/g w.w.

Due to natural variability related to intrinsic and abiotic factors, the linkage of MTs to 276 metal exposure is generally difficult to establish during field studies, even for samples from 277 278 metal-polluted environments (Legras et al., 2000; Mouneyrac et al., 2001). Accordingly, in the present study, seasonal patterns of MTs content were generally inconsistent and differed 279 between smaller and larger lobsters at both sites, while gender related differences were less 280 pronounced. The seasonality of MTs digestive gland content was previously reported by 281 282 Giarratano et al. (2016), for crabs that displayed higher MTs content in autumn than in spring. 283 Besides, Chiodi Boudet et al. (2013) reported different patterns of seasonal variations for MTs content of white shrimp Palaemonetes argentines from polluted and unpolluted marine site. 284 Individual metal concentrations and ROS in the digestive gland of N. norvegicus from both 285 sampling sites generally displayed relatively large fluctuations between body size groups and 286 287 with respect to seasons and genders. Furthermore, spatial variations were also found, suggesting different bioavailability of metals at two sites and possibly an influence of local environmental 288 conditions. 289

Seasonal variations and gender dependent differences of Cd and other metals that exert 290 291 high MTs binding affinity, were already reported for N. norvegicus digestive gland (Canli and Furness, 1993a). Nevertheless, MTs content for larger body size lobsters displayed weak 292 positive correlation (r_s=0.25) to Cd that, according to previous reports, predominantly 293 accumulates in digestive gland tissue of N. norvegicus (Canli and Furness, 1993b). The notion 294 of elevated MTs content in response to Cd could be supported by laboratory studies 295 demonstrating Cd as an effective inducer of MTs synthesis in *N. norvegicus* (Canli et al., 1997) 296 and other crustaceans (Moltó et al., 2005; Pan and Zhang, 2006). However, considering the 297 fairly weak correlation, it is difficult to speculate to what extent the level of Cd in N. norvegicus 298 digestive gland might be related to MTs content fluctuations in this particular case. According 299 to recent data on sediment metal concentrations (Cukrov et al., 2011, 2014; Ilijanić et al., 2014), 300 there are no indications of particular sediment Cd enrichment at sampling sites in comparison 301 to other offshore areas of unpolluted Adriatic and Mediterranean regions. Besides, 302 concentrations of Cd for digestive gland were in line with data previously reported for N. 303 304 norvegicus (Canli and Furness, 1993a) and lobsters H. gammarus and H. americanus (Barrento et al., 2009; Leblanc and Prince, 2012) from unpolluted sites of Atlantic coast. Similarly, low 305 306 enrichment of off-shore sediments and generally decreasing contamination trend for Cu and Zn in Kvarner Bay (Cukrov et al., 2011, 2014) could explain the lack of correlation between these 307

metals and MTs content in *N. norvegicus*. The essential metals like Cu and Zn also possess high
MTs binding ability and MTs induction potential when accumulated in excess (Amiard et al.,
2006), but it seems that a pool of MTs that bind these metals could be considered as storage
and donor for metalloproteins, such as for apohemocyanin and carboanhydrase (Henry et al.,
2012).

313 The experimental evidences on MTs-inductive potential of other toxic metals such as Hg and As are generally scarce for crustaceans (Barka et al., 2001; Barka, 2007; Vellinger et al., 314 2013), but when these organisms were used for studying the adverse effect of anthropogenic 315 contaminants in the field revealed correlations of both metals with MTs content (Faria et al., 316 2010, Martín-Díaz et al., 2009). A mild positive correlation of MTs was detected in the present 317 318 study for Hg ($r_s=0.31$) and As ($r_s=0.37$) but again only for larger lobsters, indicating that for this body size class the level of Hg and As accumulated in digestive gland tissue could be 319 sufficient to surpass the necessary threshold for MTs induction. This is consistent with the 320 essential physiological role of MTs in detoxification and storage of metals in the form of 321 322 insoluble MTs complex. Noteworthy, values for Hg concentrations exceeded those previously reported for digestive gland of N. norvegicus (Canli and Furness, 1993a) and other crustaceans 323 from the Atlantic (Barrento et al., 2009) and the Pacific (Frías-Espericueta et al., 2016). 324 Moreover, data reported herein are in agreement with consistently high Hg concentrations in 325 326 the soft tissues of *N. norvegicus* and other organisms from Mediterranean waters that, as widely 327 emphasised before, could be related to the large cinnabar deposits in the Mediterranean basin (up to 55% of total world reserves) and slow turnover of Mediterranean waters through the 328 Gibraltar strait (Perugini et al., 2009; Renzoni et al., 1998). Besides, Hg enrichment of 329 sediments and biota in the Northern Adriatic is predominantly linked to continuous discharge 330 331 from nearby industrialized zones and large Hg mining sites (Kotnik et al., 2015). Levels of As were higher than in digestive gland of crustaceans from the Atlantic (Barrento et al., 2009; 332 Leblanc and Prince, 2012) and the Pacific (Lewtas et al., 2014; Metian et al., 2010). A relatively 333 high amount of total As was already reported for N. norvegicus digestive gland from the same 334 Adriatic area (Sekulić et al., 1993). The findings of high As levels in comparison to crustaceans 335 from other geographical areas are consistent with previous reports for temperate Mediterranean 336 and tropical Caribbean seas (Fattorini et al., 2006) and could be linked to the influence of 337 environmental conditions, particularly temperature and salinity fluctuations (Valentino-Álvarez 338 339 et al., 2013; Vellinger et al., 2012).

In contrast to larger organisms, the lack or even inverse relationship of Cd, Hg and As accumulation and MTs content were found for small lobsters. The discrepancy between smaller and larger organisms could be explained by possibly major influence of body size on metal detoxification efficiency in *N. norvegicus*. This hypothesis needs further experimental verification.

Previous laboratory and field studies demonstrated positive correlation of Pb, Cr and Mn to MTs content in some crustaceans (Lavadras et al., 2014; Martín-Díaz et al., 2009). On the other hand, at exposure concentrations of Pb relevant for contaminated marine environment, the accumulation of Pb in digestive gland tissue of shrimp *L. vannamei* was not observed and could not be linked to MTs content increase (Nunez-Nogueira et al., 2010). Hence, the lack of correlation to MTs could be explained by relatively low concentrations of Pb that were two to tenfold lower than values previously reported by Canli and Furness (1993a) for digestive gland

of N. norvegicus, and within values for other crustaceans from low to moderately contaminated 352 marine areas (Canli et al., 2001; Leblanc and Prince, 2012; Lewtas et al., 2014; Yilmaz and 353 Yilmaz, 2007). Similarly, concentrations of Cr reported herein were comparable to those 354 reported for shrimps from unpolluted aquaculture site in the Pacific (Metian et al., 2010), 355 although some studies reported lower levels for other decapode crustaceans collected from sites 356 357 of varying contamination degree (Ciftci et al 2011; Leblanc and Prince, 2012; Pereira et al 358 2009). It is also important to consider that both metals (Pb in particular) accumulate more effectively in other tissues of N. norvegicus, such as the gills (Cenov, 2017) further limiting the 359 possibility for linking the observed fluctuations of these metals to MTs content in digestive 360 gland. Accumulation of Mn was higher than previously found for digestive gland of lobsters N. 361 norvegicus (Baden et al., 1999) and *H. americanus* from low to moderately contaminated areas 362 in the Atlantic (Leblanc and Prince, 2012) but generally lower than in blue shrimp Litopenaeus 363 stylirostris from the Pacific (Metian et al., 2010). The lack of correlation between Mn and MTs 364 content is consistent with the tendency of slow Mn accumulation in the digestive gland with 365 366 respect to gills or exoskeleton in particular (Cenov, 2017), and the variable pattern of accumulation recorded for N. norvegicus might reflect dietary intake Mn rather than its level in 367 the surrounding environment (Ericsson and Baden, 1998). 368

As mentioned above, some metals accumulate more effectively in the gills than in the digestive gland. Thus, the response of MTs in the gills to metal accumulation should be also considered for investigation, taking also into account that relatively high MTs content detected in this particular tissue of *N. norvegicus* (Canli et al., 1993).

While Cd is mostly distributed in soluble cytosol (Pedersen et al., 2014) some metals are mainly detoxified as insoluble metal-rich granules, as reported for some crustaceans (Barka, 2007; Legras et al., 2000; Mouneyrac et al., 2001; Nunez – Noguiera et al., 2010). Since the total concentration of metals was taken into account here, their potentially toxic effect reflected in elevated MTs content might have remained obscured. Clearly, information on partitioning of metals between soluble and insoluble fraction is needed to further explain the MTs content fluctuations trends in relation to metal accumulation in *N. norvegicus*.

- Levels of ROS displayed different seasonal pattern for small and large lobsters possibly in 380 relation to fluctuations of environmental abiotic and biotic factors within the investigated areas, 381 that were shown to influence the balance between pro-oxidant and antioxidant activity and 382 383 maintenance of a steady-state ROS level in crustaceans (Liu et al., 2007; Schvezov et al., 2015). A protective role of MTs acting as scavengers of ROS arising from the action of metals was 384 reported by Moltó et al (2007). Furthermore, Felix-Portillo et al (2014) reported the increased 385 MTs mRNA expression following hypoxia exposure of white shrimps L. vannamei suggesting 386 the possible role of MTs in the ROS - detoxifying mechanism. Another experimental study 387 showed an increase of MTs level following exposure of L. vannamei to Cd, and a concomitant 388 decrease of ROS production (Lobato et al., 2013). In this respect, a modest negative relationship 389 between MTs and ROS accumulation ($r_s = -0.45$) for small but not for large organisms, indicates 390 a body size dependent capacity of MTs to counteract the oxidative radicals. However, data on 391 392 antioxidative system components and in particular on lipid peroxidation are needed for a more detailed picture on the capacity of *N. norvegicus* to cope with potential pro oxidants. 393
- A gender-related specificity of MTs content in digestive gland of lobsters was not detected in the current study. Our findings are opposite to recent evidences of gender dependent MTs

content variations reported for crabs Neohelice granulata (Buzzi and Marcovecchio, 2016) and 396 Callinectes sp. (Lavradas et al., 2014). Moreover, the use of only one gender was suggested for 397 investigations of metals accumulation and biochemical responses in the tissues of crustaceans 398 (Giarratano et al., 2016; Martín-Díaz et al., 2009). The absence of gender dependent Cd 399 concentrations clearly contrasted previous findings of significantly higher level of this metal in 400 the digestive gland of N. norvegicus females (Canli and Furness, 1993a). Conversely, a notable 401 402 and consistently higher accumulation in males was displayed for As and Hg for both body-size groups and in both seasons, in accordance to previous study on N. norvegicus (Canli and 403 Furness, 1993a). The observed differences may be due to larger dimensions of males than 404 females sampled within the frame of the present study. In fact, Barrento et al (2009) found 405 406 higher As concentration in digestive gland of females that in that particular survey displayed 407 faster growth rate.

Variations in energy reserves, in particular lipids content, indicated that the physiology of 408 N. norvegicus was influenced by season in both small and large organisms. Lipids content 409 410 variation could be related to season dependent N. norvegicus feeding activity that in the Adriatic Sea tends to be higher in autumn (Cristo and Cartes, 1998). Generally lower level of lipids in 411 spring could be a consequence of reduced feeding rhythm during the winter (Watts et al., 2016). 412 More expressed consumption of energy reserves is displayed by males that commonly undergo 413 moults more frequently than females (Sardà, 1995). As suggested by Rosa and Nunes (2002), 414 415 lipids stored in digestive gland tissue could be more important for moulting activity than for oogenesis, which seems to depend on dietary intake of lipids. Mostly moderate negative 416 correlations of MTs content and metals accumulation with lipids in particular, observed here 417 for both body size categories, prompts for caution when interpreting these parameters in N. 418 419 norvegicus, taking into consideration the expressed seasonal fluctuations on biochemical composition in digestive gland. For improved interpretation of chemical and biological data in 420 relation to physiological conditions of N. norvegicus, the digestive gland glycogen level data 421 would be also helpful, since it was shown to decline during starvation (Philp et al., 2015). 422

423 It is important to note that data interpretation could be impaired by confounding factors such as the reproductive status that affects metal accumulation and MTs content in some 424 crustacean species (Mouneyrac et al., 2001). In the present study, the influence of reproductive 425 cycle on fluctuations of MTs level and metal concentrations in N. norvegicus digestive gland 426 observed could not be tackled, due to low proportion of females in trawl catch with respect to 427 males. It is generally accepted that the reproductive season of N. norvegicus in the Adriatic Sea 428 429 peaks in late spring and summer while the proportion of mature females declines in autumn, in accordance to changes in maturation stages of the ovaries (Orsi-Rellini et al., 1998). 430 Nevertheless, the possible linkage of observed MTs content and metal accumulation seasonality 431 and assumed differences in the reproductive stage between N. norvegicus sampled in spring and 432 in autumn was not discerned in the present study, possibly due to expressed spatial and temporal 433 heterogeneity of individuals in terms of gonad maturation stage even within the same body size 434 group. This was suggested by occasional and unsynchronised occurrence of larger eggs-435 carrying females in both seasons at both sites, and only smaller eggs-carrying females in autumn 436 at S1, but not at site S2. Thus, the important issue of reproductive cycle interference with MTs 437 438 content and metal accumulation in N. norvegicus still remains unresolved.

An earlier study suggested that metal accumulation in the digestive gland tissue of N. 439 norvegicus may vary considerably over different moulting stages (Canli and Furness, 1993a) 440 that were not assessed in the present study. Similar observations were also reported for other 441 crustaceans (Brouwer et al., 1992; Nørum et al., 2005). It is generally accepted that moulting 442 frequency of adult N. norvegicus decreases with age and differs between females and males, 443 444 being, as already mentioned, more frequent in the latter (Sardà, 1995). Whether factors such as 445 moulting stages could be responsible for MTs content variation in the tissues of crustaceans is currently not sufficiently clear. Considering relatively heterogeneous carapace length of 446 samples and obvious differences in the MTs trends between two body size groups, it is plausible 447 that moulting frequency and concomitant, possibly gender-dependent size increments of N. 448 449 norvegicus (Sardà, 1995) could represent an additional confounding factor for the interpretation of MTs content changes, in particular in relation to metals accumulation in the digestive gland 450 tissue. 451

Finally, the comparison of MTs content between N. norvegicus digestive gland from the 452 453 Kvarner Bay (present study) and that of other crustacean species from worldwide coastal and off-shore areas is impaired due to the well-known discrepancies between MTs content data 454 related to differences in the method for MTs quantification (Pedersen et al., 2008). Considering 455 the results previously obtained by spectrophotometric sulphydryl method (Viarengo et al., 456 1997), the values for MTs content presented herein are of the same order of magnitude as that 457 of crab *Neohelice granulata* (Buzzi and Marcovecchio, 2016). Values higher by approximately 458 one order of magnitude were reported for blue crab Callinectes sapidus from unpolluted sites 459 (De Martinez Gaspar Martins and Bianchini, 2009). 460

A definition of threshold for the background is recommended to facilitate the interpretation 461 462 of biological response, but requires the synthesis of field data both from uncontaminated and contaminated environments (Davies and Vethaak, 2012). The results of this study represent the 463 only relatively comprehensive data available on the MTs content so far for field sampled N. 464 norvegicus from the Mediterranean Sea. Furthermore, the significance of MTs content value 465 deviation from the threshold value is currently not sufficiently understood for this benthic 466 crustacean species. Thus, the threshold value for background level (mean + 1σ , 25.2 nmol/g 467 w.w.) could be considered only as a tentative suggestion. Obviously, further investigation and 468 collection of more data, particularly from metal enriched gradients are required to establish 469 regionally specific threshold values crucial for discerning the natural variability of MTs content 470 from potential adverse effect of toxic metals in *N. norvegicus*. 471

This study presents the first report on the metallothioneins content in the digestive gland 472 of N. norvegicus taking into account the effect of body size, gender and spatio - temporal 473 variations. Body size had significant influence on MTs content variations that displayed 474 generally inconsistent seasonal patterns, raising questions whether and to what extent the effect 475 of metals exposure could be masked and remain undetected. By contrast, differences between 476 males and females were negligible. Despite seasonal fluctuations of both MTs content and metal 477 accumulation, the observed mild positive relationship with Cd, Hg and As and was in 478 479 accordance to the well-known metal-scavenging function of these proteins. A negative relationship with ROS reinforced the notion of possible MTs involvement in antioxidative 480 response. This study also stressed that variations of biochemical and chemical parameters 481 measured in *N. norvegicus* digestive gland tissue could be linked to energy reserves, particularly 482

483	lipids. Thus, data presented here provide a solid starting point for future studies that should be simed in particular to filling the knowledge gaps concerning MTs response to increased metal
484	aimed in particular to filling the knowledge gaps concerning MTs response to increased metal
485	body burden.
486	
487	Acknowledgements
488	This research was supported by Teaching Institute of Public Health of Primorje-Gorski
489	Kotar County.
490	
491	References
492	
493	Amiard, JC., Amiard-Triquet, C., Barka, S., Pellerin, J. & Rainbow, P.S., 2006.
494	Metallothioneins in aquatic invertebrates: Their role in metal detoxification and their use as
495	biomarkers. Aquat. Toxicol. 76 (2), 160-202.
496	
497	Baden, S.P., Eriksson, S.P. & Gerhardt, L., 1999. Accumulation and elimination kinetics
498	of manganese from different tissues of the Norway lobster Nephrops norvegicus (L.). Aquat.
499	Toxicol. 46 (2), 127-137
500	
501	Barka, S., Pavillon, JF. & Amiard, JC., 2001. Influence of different essential and non-
502	essential metals on MTLP levels in the Copepod Tigriopus brevicornis. Comp. Biochem.
503	Physiol. C Toxicol. Pharmacol. 128 (4), 479-493.
504	
505	Barka S., 2007. Insoluble detoxification of trace metals in a marine copepod Tigriopus
506	brevicornis (Muller) exposed to copper, zinc, nickel, cadmium, silver and mercury.
507	Ecotoxicology 16 491-502.
508	Leotoxicology 10 471-502.
	Barrento, S., Marques, A., Teixeira, B., Carvalho, M.L., Vaz-Pires, P. & Nunes, M.L.,
509	
510	2009. Accumulation of elements (S, As, Br, Sr, Cd, Hg, Pb) in two populations of <i>Cancer</i>
511	pagurus: Ecological implications to human consumption. Food Chem. Toxicol. 47 (1), 150-
512	156.
513	
514	Barrick, A., Châtel, A., Marion, JM., Perrein-Ettajani, H., Bruneau, M. & Mouneyrac,
515	C., 2016. A novel methodology for the determination of biomarker baseline levels in the marine
516	polychaete Hediste diversicolor. Mar. Pollut. Bull. 108 (1-2), 275-280.
517	
518	Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram
519	quantities of protein utilizing the principle of protein-dye binding. Anal. Biochem. 72, 248-254.
520	
521	Brouwer, M., Engel, D. W., Bonaventura, J. & Johnson, G. A., 1992. In vivo magnetic
522	resonance imaging of the blue crab Callinectes sapidus: effect of cadmium accumulation in
523	tissues on proton relaxation properties. J. Exp. Zool. 261, 32–40.
524	

525	Buzzi, N.S. & Marcovecchio, J.E., 2016. A baseline study of the metallothioneins
526	induction and its reversibility in Neohelice granulata from the Bahía Blanca Estuary
527	(Argentina). Mar. Pollut. Bull. 112 (1-2), 452-458.
528	
529	Canli, M. & Furness, R.W., 1993a. Heavy Metals in Tissues of the Norway Lobster
530	Nephrops norvegicus: Effects of Sex, Size and Season. Chem. Ecol. 8 (1), 19-32.
531	
532	Canli, M., Furness, R.W., 1993b. Toxicity of heavy metals dissolved in sea water and
533	influences of sex and size on metal accumulation and tissue distribution in the Norway lobster
534	Nephrops norvegicus. Mar. Environ. Res. 36 (4), 217-236.
535	
536	Canli, M., Stagg, R.M. & Rodger, G., 1997. The induction of metallothionein in tissues of
537	the Norway lobster Nephrops norvegicus following exposure to cadmium, copper and zinc: The
538	relationships between metallothionein and the metals. Environ. Pollut. 96 (3), 343-350.
539	
540	Canli, M., Kalay, M. & Ay, Ö., 2001. Metal (Cd, Pb, Cu, Zn, Fe, Cr, Ni) concentrations in
541	tissues of a fish Sardina pilchardus and a prawn Peaenus japonicus from three stations on the
542	Mediterranean Sea. Bull. Environ. Contam. Toxicol. 67 (1), 75-82.
543	
544	Cenov, A., 2017. Biološki odgovor prirodnih populacija škampa Nephrops norvegicus
545	(Linnaeus, 1758) u sjevernom jadranu na toksične metale [Biological response of natural
546	populations of the lobster Nephrops norvegicus (Linnaeus, 1758), in the northern Adriatic sea
547	to toxic metals]. PhD thesis, University of Zagreb, Zagreb, Croatia, pg 131.
548	
549	Chiodi Boudet, L., Polizzi, P., Romero, M.B., Robles, A. & Gerpe, M., 2013. Lethal and
550	sublethal effects of cadmium in the white shrimp Palaemonetes argentinus: A comparison
551	between populations from contaminated and reference sites. Ecotoxicol. Environ. Safe. 89, 52-
552	58.
553	
554	Çiftçi, N., Cicik, B., Erdem, C., Ay, Ö., Karayakar, F. & Karaytug, S., 2011. Accumulation
555	of chromium in hepatopancreas, gill and muscle tissues of Callinectes sapidus. Fresenius
556	Environ. Bull. 20 (4A), 1089-1092.
557	
558	Cristo, M. & Cartes, J.E., 1998. A comparative study of the feeding ecology of Nephrops
559	norvegicus (L.), (Decapoda: Nephropidae) in the bathyal Mediterranean and the adjacent
560	Atlantic. Sci. Mar. 62 (SUPPL.1), 81-90.
561	
562	Cukrov, N., Frančišković-Bilinski, S., Hlača, B. & Barišić, D., 2011. A recent history of
563	metal accumulation in the sediments of Rijeka harbor, Adriatic Sea, Croatia. Mar. Pollut. Bull.
564	62 (1), 154-167.
565	
566	Cukrov, N., Frančišković-Bilinski, S. & Bogner, D., 2014. Metal contamination recorded
567	in the sediment of the semi-closed Bakar Bay (Croatia). Environ. Geochem. Health 36 (2), 195-
568	208.

569	
570	Davies, I.M. & Vethaak, A.D., 2012. Integrated marine environmental monitoring of
571	chemicals and their effects. ICES Cooperative Research Report No. 315.
572	
573	De Martinez Gaspar Martins, C. & Bianchini, A., 2009. Metallothionein-like proteins in
574	the blue crab Callinectes sapidus: Effect of water salinity and ions. Comp. Biochem. Physiol.
575	A Mol. Integr. Physiol. 152 (3), 366-371.
576	
577	Eriksson, S.P. & Baden, S.P., 1998. Manganese in the haemolymph and tissues of the
578	Norway lobster, Nephrops norvegicus (L.), along the Swedish west coast, 1993-1995.
579	Hydrobiologia 375-376, 255-264.
580	
581	Faria, M., Huertas, D., Soto, D.X., Grimalt, J.O., Catalan, J., Riva, M.C. & Barata, C.,
582	2010. Contaminant accumulation and multi-biomarker responses in field collected zebra
583	mussels (Dreissena polymorpha) and crayfish (Procambarus clarkii), to evaluate toxicological
584	effects of industrial hazardous dumps in the Ebro river (NE Spain). Chemosphere 78 (3), 232-
585	240.
586	
587	Fattorini, D., Notti, A. & Regoli, F., 2006. Characterization of arsenic content in marine
588	organisms from temperate, tropical, and polar environments. Chem. Ecol. 22 (5), 405-414.
589	
590	Felix-Portillo, M., Martinez-Quintana, J.A., Peregrino-Uriarte, A.B. & Yepiz-Plascencia,
591	G., 2014. The metallothionein gene from the white shrimp Litopenaeus vannamei:
592	Characterization and expression in response to hypoxia. Mar. Environ. Res. 101 (1), 91-100.
593	
594	Frías-Espericueta, M.G., Ramos-Magaña, B.Y., Ruelas-Inzunza, J., Soto-Jiménez, M.F.,
595	Escobar-Sánchez, O., Aguilar-Juárez, M., Izaguirre-Fierro, G., Osuna-Martínez, C.C. &
596	Voltolina, D., 2016. Mercury and selenium concentrations in marine shrimps of NW Mexico:
597	health risk assessment. Environ. Monit. Assess. 188 (11), art. no. 629,
598	
599	Giarratano, E., Gil, M.N., Marinho, C.H. & Malanga, G., 2016. Metals from mine waste
600	as potential cause of oxidative stress in burrowing crab <i>Neohelice granulata</i> from San Antonio
601	bay. Ecotoxicol. Environ. Safe. 132, 68-76.
602	•
603	Glad, M., 2017. Utjecaj veličine i spola na raspodjelu esencijalnih elemenata u tkivima
604	škampa Nephrops norvegicus (Linnaeus, 1758) [The effect of size and sex on distribution of
605	essential elements in tissue of Norway lobster Nephrops norvegicus (Linnaeus, 1758)] PhD
606	thesis, University of Zagreb, Zagreb, Croatia, pg 153.
607	
608	Henry, R.P., Lucu, Č., Onken, H. & Weihrauch, D., 2012. Multiple functions of the
609	crustacean gill: Osmotic/ionic regulation, acid-base balance, ammonia excretion, and
610	bioaccumulation of toxic metals. Front. Physiol. 3 NOV, art. no. Article 431,.
611	• • • • • • • • •

612	Ilijanić, N., Miko, S., Petrinec, B. & Franić, Z., 2014. Metal deposition in deep sediments
613	from the Central and South Adriatic Sea. Geol. Croat. 67 (3), 185-205.
614	
615	Kotnik, J., Horvat, M., Ogrinc, N., Fajon, V., Žagar, D., Cossa, D., Sprovieri, F. & Pirrone,
616	N., 2015. Mercury speciation in the Adriatic Sea. Mar. Pollut. Bull. 96 (1-2), 136-148.
617	
618	Lavradas, R.T., Hauser-Davis, R.A., Lavandier, R.C., Rocha, R.C.C., Saint' Pierre, T.D.,
619	Seixas, T., Kehrig, H.A. & Moreira, I., 2014. Metal, metallothionein and glutathione levels in
620	blue crab (Callinectes sp.) specimens from southeastern Brazil. Ecotoxicol. Environ. Safe. 107,
621	55-60.
622	
623	Leblanc, L.A. & Prince, D., 2012. Metal concentrations in tissues of American lobsters
624	(Homarus americanus, Milne-Edwards) with epizootic shell disease. J. Shellfish Res. 31 (2),
625	543-553.
626	
627	Legras, S., Mouneyrac, C., Amiard, J.C., Amiard-Triquet, C. & Rainbow, P.S., 2000.
628	Changes in metallothionein concentrations in response to variation in natural factors (salinity,
629	sex, weight) and metal contamination in crabs from a metal-rich estuary. J. Exp. Mar. Bio. Ecol.
630	246 (2), 259-279.
631	
632	Lewtas, K.L.M., Birch, G.F. & Foster-Thorpe, C., 2014. Metal accumulation in the
633	greentail prawn, Metapenaeus bennettae, in Sydney and Port Hacking estuaries, Australia.
634	Environ. Sci. Pollut. Res. 21 (1) 704-716.
635	
636	Liu, Y., Wang, WN., Wang, AL., Wang, JM. & Sun, RY., 2007. Effects of dietary
637	vitamin E supplementation on antioxidant enzyme activities in Litopenaeus vannamei (Boone,
638	1931) exposed to acute salinity changes. Aquaculture 265 (1-4), 351-358.
639	
640	Lobato, R.O., Nunes, S.M., Wasielesky, W., Fattorini, D., Regoli, F., Monserrat, J.M. &
641	Ventura-Lima, J., 2013. The role of lipoic acid in the protection against of metallic pollutant
642	effects in the shrimp Litopenaeus vannamei (Crustacea, Decapoda). Comp. Biochem. Physiol.
643	A Mol. Integr. Physiol. 165 (4), 491-497.
644	
645	Maria, V.L., Santos, M.A. & Bebianno, M.J., 2009. Contaminant effects in shore crabs
646	(Carcinus maenas) from Ria Formosa Lagoon. Comp. Biochem. Physiol. C Toxicol.
647	Pharmacol. 150 (2), 196-208.
648	
649	Martín-Díaz, M.L., Blasco, J., Sales, D. & DelValls, T.A., 2009. The use of a kinetic
650	biomarker approach for in situ monitoring of littoral sediments using the crab Carcinus maenas.
651	Mar. Environ. Res. 68 (2), 82-88.
652	
653	Metian, M., Hédouin, L., Eltayeb, M.M., Lacoue-Labarthe, T., Teyssié, JL., Mugnier, C.,
654	Bustamante, P. & Warnau, M., 2010. Metal and metalloid bioaccumulation in the Pacific blue

- shrimp *Litopenaeus stylirostris* (Stimpson) from New Caledonia: Laboratory and field studies.
 Mar. Pollut. Bull. 61 (7-12), 576-584.
- 657

Moltó, E., Bonzón-Kulichenko, E., Del Arco, A., López-Alañón, D.M., Carrillo, O.,
Gallardo, N. & Andrés, A., 2005. Cloning, tissue expression and metal inducibility of an
ubiquitous metallothionein from *Panulirus argus*. Gene 361 (1-2), 140-148.

661

Moltó, E., Bonzón-Kulichenko, E., Gallardo, N. & Andrés, A., 2007. MTPA: A crustacean
metallothionein that affects hepatopancreatic mitochondrial functions. Arch. Biochem.
Biophys. 467 (1), 31-40.

665

Mouneyrac, C., Amiard-Triquet, C., Amiard, J.C. & Rainbow, P.S., 2001. Comparison of
metallothionein concentrations and tissue distribution of trace metals in crabs (*Pachygrapsus marmoratus*) from a metal-rich estuary, in and out of the reproductive season. Comp. Biochem.
Physiol. C Toxicol. Pharmacol. 129 (3), 193-209.

670

Nunez-Nogueira, G., Mouneyrac, C., Muntz, A. & Fernandez-Bringas, L., 2010.
Metallothionein-like proteins and energy reserve levels after Ni and Pb exposure in the pacific
white prawn *Penaeus vannamei*. J. Toxicol., art. no. 407360,

674

Nørum, U., Bondgaard, M., Pedersen, T.V. & Bjerregaard, P., 2005. *In vivo* and *in vitro*cadmium accumulation during the moult cycle of the male shore crab *Carcinus maenas* Interaction with calcium metabolism. Aquat. Toxicol. 72 (1-2 SPEC. ISS.), 29-44.

Orsi-Relini L., Zamboni, A., Fiorentino, F. & Massi, D., 1998. Reproductive patterns in
Norway lobster *Nephrops norvegicus* (L.), (Crustacea Decapoda Nephropidae) of different
Mediterranean areas. Sci. Mar. 62 (SUPPL.1), 25-41.

Ortega, P., Vitorino, H.A., Moreira, R.G., Pinheiro, M.A.A., Almeida, A.A., Custódio,
M.R. & Zanotto, F.P., 2017. Physiological differences in the crab *Ucides cordatus* from two
populations inhabiting mangroves with different levels of cadmium contamination. Environ.
Toxicol. Chem. 36 (2), 361-371.

687

682

Pan, L. & Zhang, H., 2006. Metallothionein, antioxidant enzymes and DNA strand breaks
as biomarkers of Cd exposure in a marine crab, *Charybdis japonica*. Comp. Biochem. Physiol.
C Toxicol. Pharmacol. 144 (1), 67-75.

691

Pedersen, K.L., Pedersen, S.N., Knudsen, J. & Bjerregaard, P., 2008. Quantification of
metallothionein by differential pulse polarography overestimates concentrations in crustaceans.
Environ. Sci. Technol. 42 (22), 8426-8432.

695

Pedersen, K.L., Bach, L.T. & Bjerregaard, P., 2014 Amount and metal composition of
midgut gland metallothionein in shore crabs (*Carcinus maenas*) after exposure to cadmium in
the food. Aquat. Toxicol. 150, 182-188.

699	
700	Pereira, P., de Pablo, H., Dulce Subida, M., Vale, C. & Pacheco, M., 2009. Biochemical
701	responses of the shore crab (Carcinus maenas) in a eutrophic and metal-contaminated coastal
702	system (Óbidos lagoon, Portugal). Ecotoxicol. Environ. Safe. 72 (5), 1471-1480.
703	
704	Perugini, M., Visciano, P., Manera, M., Zaccaroni, A., Olivieri, V. & Amorena, M., 2009.
705	Levels of total mercury in marine organisms from Adriatic Sea, Italy. Bull. Environ. Contam.
706	Toxicol. 83 (2), 244-248.
707	
708	Philp, H., Albalat, A. & Marteinsdóttir, G., 2015. Live holding of Nephrops norvegicus
709	(Linnaeus, 1758) in land-based facilities: Health and condition effects. Mar. Biol. Res. 11 (6),
710	603-612.
711	
712	Piccinetti, C., Vrgoč, N., Marčeta, B. & Manfredi, C., 2012. Recent state of demersal
713	resources in the Adriatic Sea. Acta Adriat. Monograph Series no.5, 220 pp.
714	
715	Renzoni, A., Zino, F. & Franchi, E., 1998. Mercury levels along the food chain and risk
716	for exposed populations. Environ. Res. 77 (2), 68-72.
717	
718	Rosa, R. & Nunes, M.L., 2002. Biochemical changes during the reproductive cycle of the
719	deep-sea decapod Nephrops norvegicus on the south coast of Portugal. Mar. Biol. 141 (6), 1001-
720	1009.
721	
722	RStudio Team, 2015. RStudio: Integrated Development for R. RStudio, Inc., Boston, MA
723	URL <u>http://www.rstudio.com/</u> .
724	
725	Sardà, F., 1995. A review (1967-1990) of some aspects of the life history of Nephrops
726	norvegicus. ICES Marine Sciencia Symposium 199., 78-88.
727	
728	Schvezov, N., Lovrich, G.A., Florentín, O. & Romero, M.C., 2015. Baseline defense
729	system of commercial male king crab Lithodes santolla from the Beagle Channel. Comp.
730	Biochem. Physiol. A Mol. Integr. Physiol. 181, 18-26.
731	
732	Sekulić, B., Sapunar, J. & Bažulić, D., 1993. Arsenic in Norway lobster (Nephrophs
733	norvegicus L.) from Kvarnerić Bay - Northeastern Adriatic. Bull. Environ. Contam. Toxicol.
734	51 (3), 460-463.
735	
736	Valentino-Álvarez, J.A., Núñez-Nogueira, G. & Fernández-Bringas, L., 2013. Acute
737	toxicity of arsenic under different temperatures and salinity conditions on the white shrimp
738	<i>Litopenaeus vannamei</i> . Biol. Trace Elem. Res. 152 (3), 350-357.
739	
740	Vellinger, C., Felten, V., Sornom, P., Rousselle, P., Beisel, JN. & Usseglio-Polatera, P.,
741	2012. Behavioural and physiological responses of <i>Gammarus pulex</i> exposed to cadmium and

arsenate at three temperatures: Individual and combined effects. PLoS ONE 7 (6), art. no. 742 e39153 743 744 Vellinger, C., Gismondi, E., Felten, V., Rousselle, P., Mehennaoui, K., Parant, M. & 745 Usseglio-Polatera, P., 2013. Single and combined effects of cadmium and arsenate in 746 Gammarus pulex (Crustacea, Amphipoda): Understanding the links between physiological and 747 748 behavioural responses. Aquat. Toxicol. 140-141, 106-116. 749 750 Viarengo, A., Ponzano, E., Dondero, F. & Fabbri, R., 1997. A simple spectrophotometric method for metallothionein evaluation in marine organisms: An application to Mediterranean 751 752 and Antarctic molluscs. Mar. Environ. Res. 44 (1), 69-84. 753 Watts, A.J.R., Albalat, A., Smith, I.P., Atkinson, R.J.A. & Neil, D.M., 2016. Seasonal 754 755 nutritional status in Norway lobsters, Nephrops norvegicus (L.): are females nutritionally 756 compromised over the winter? Mar. Biol. Res. 12 (6), 563-572. 757 758 Yilmaz, A.B. & Yilmaz, L., 2007. Influences of sex and seasons on levels of heavy metals in tissues of green tiger shrimp (Penaeus semisulcatus de Hann, 1844). Food Chem. 101 (4), 759 1664-1669. 760 761 Zhu, Z., DeRose, E.F., Mullen, G.P., Petering, D.H. & Shaw III, C.F., 1994. Sequential 762 proton resonance assignments and metal cluster topology of lobster metallothionein-1. 763 Biochemistry 33 (30), 8858-8865. 764 765

18

Figure captions

Fig 1. Map of sampling area in the Kvarner Bay, NE Adriatic Sea.

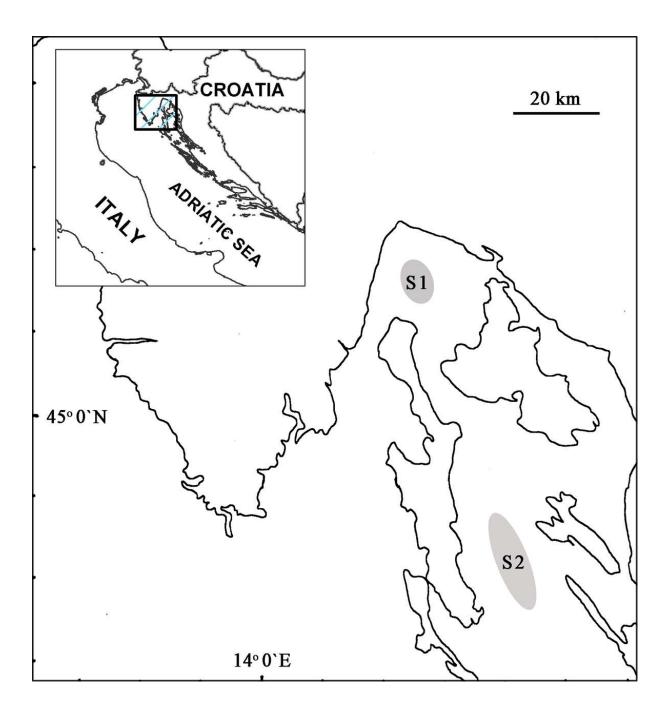
Fig 2. MTs content (nmol/g w.w.) in the digestive gland of small (<36mm) and large (>36 mm) *Nephrops norvegicus* from sites S1 and S2 in autumn (Aut) and spring (Spr). Square boxes indicate lower and upper quartile and whiskers represent minimum and maximum data values (1.5 interquartile range). Medians are depicted by solid line, and outliers as circles. \Box – females; [#]p<0.05 - significant differences between seasons

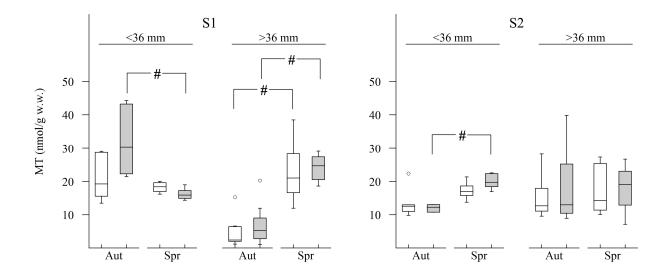
Fig 3. Concentrations of Cd, As, Hg, Pb, Cr, Mn (μ g/g d.w.) in the digestive gland of small (<36mm) and large (>36 mm) *Nephrops norvegicus* from sites S1 and S2 in autumn (Aut) and spring (Spr). Square boxes indicate lower and upper quartile and whiskers represent minimum and maximum data values (1.5 interquartile range). Medians are depicted by solid line, and outliers as circles. \Box – females; \blacksquare – males; *p<0.05 - significant difference between males and females; [#]p<0.05 - significant differences between seasons

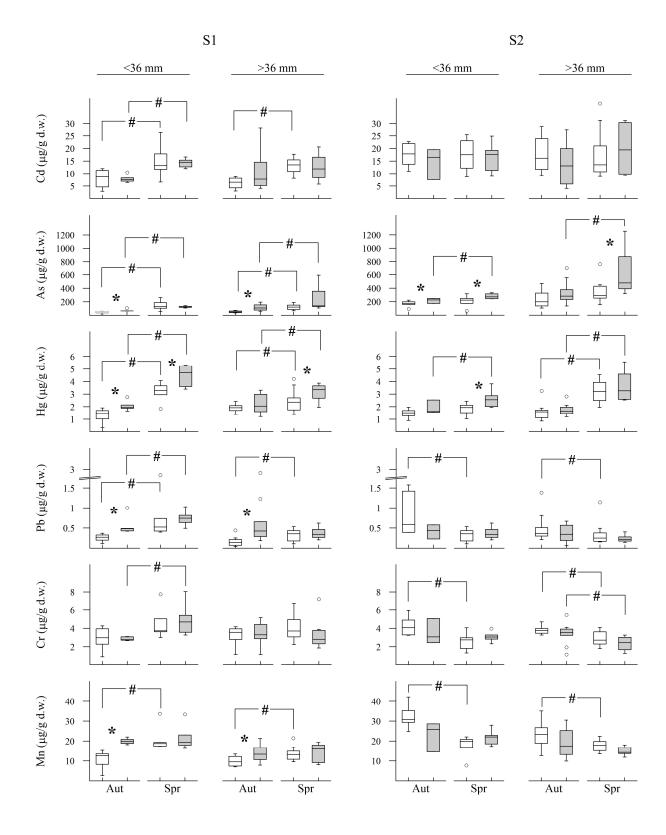
Fig 4. Content of ROS (mmol/mg w.w) in the digestive gland of small (<36mm) and large (>36 mm) *Nephrops norvegicus* from sites S1 and S2 in autumn (Aut) and spring (Spr). \Box – females; Square boxes indicate lower and upper quartile and whiskers represent minimum and maximum data values (1.5 interquartile range). Medians are depicted by solid line, and outliers as circles. \Box – females; \blacksquare – males; *p<0.05 - significant difference between males and females; #p<0.05 - significant differences between seasons

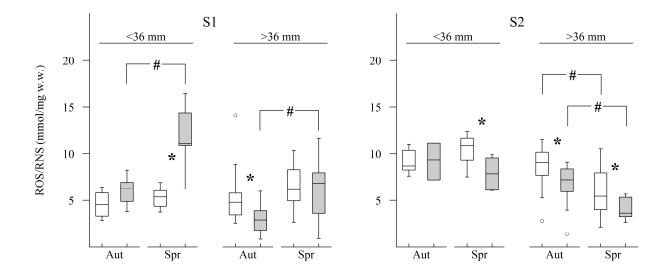
Fig 5. Score plots and variable loadings plots of principal component analysis (PCA) based on MTs content, concentrations of Cd, As, Hg, Pb, Cr, Mn, Cu and Zn, ROS content, lipids and proteins concentration in the digestive gland of small (<36 mm, upper panel) and large (>36 mm, lower panel) male and female lobsters from sites S1 and S2. Each point corresponds to one individual score. Data for Cu and Zn were provided by Glad (personal communication).

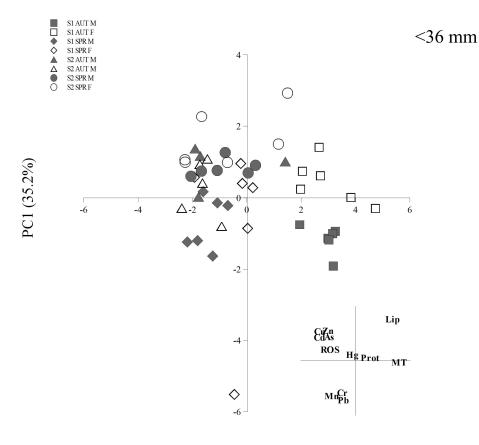
Fig 6. Frequency histogram and Gaussian distribution of values for MTs content in digestive gland of 144 *Nephrops norvegicus* samples in total.



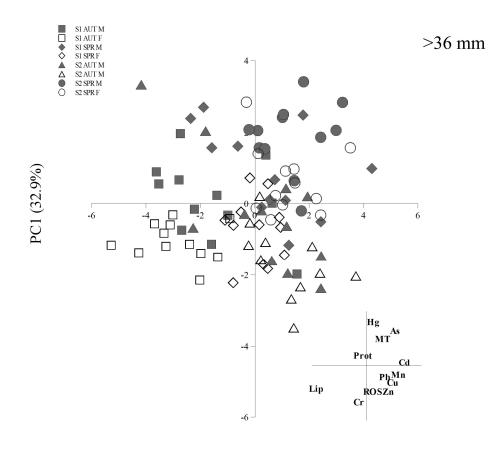








PC2 (24%)



PC2 (18.7%)

