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Source / Izvornik: **Data in Brief, 2017, 15, 738 - 741**

Journal article, Published version

Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

<https://doi.org/10.1016/j.dib.2017.09.060>

Permanent link / Trajna poveznica: <https://um.nsk.hr/um:nbn:hr:184:455911>

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Download date / Datum preuzimanja: **2025-03-10**



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Data in Brief

journal homepage: www.elsevier.com/locate/dib

Data Article

Data set of proteomic analysis of food borne pathogens after treatment with the disinfectants based on pyridoxal oxime derivatives

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ARTICLE INFO

Article history:

Received 7 June 2017

Accepted 26 September 2017

Available online 29 September 2017

ABSTRACT

Food borne pathogens, namely the Gram-positive bacterium *Bacillus subtilis* and the Gram-negative bacterium *Escherichia coli*, were grown under the inhibition with four different disinfectants based on chloride and bromide salts of pyridinium oxime. Bacterial samples were subjected to the sequential extraction of proteins and the in-solution tryptic digestion of obtained extracts was performed prior to the identification of proteins with LC-ESI-MS/MS. Proteomic analysis identified up- and down-regulated proteins in these bacteria after treatment with each compound. The tables with differently expressed proteins are presented with this article.

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DOI of original article: <http://dx.doi.org/10.1016/j.foodres.2017.06.016>

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<http://dx.doi.org/10.1016/j.dib.2017.09.060>

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Specifications Table

Subject area	Biochemistry
More specific subject area	Proteomics of foodborne bacteria
Type of data	Tables, Excel files, Word documents
How data was acquired	Tandem mass spectrometry (LC-MS/MS) using LTQ Velos Orbitrap mass spectrometer (Thermo Electron Corp., San Jose, CA, USA)
Data format	Filtered and raw
Experimental factors	Bacterial cultures of Gram-positive bacterium <i>Bacillus subtilis</i> and Gram-negative bacterium <i>Escherichia coli</i> were treated with four different ammonium salts of pyridinium oxime
Experimental features	Sequential extraction of proteins from bacterial samples was performed using ReadyPrep Extraction kit (BioRad, Hercules, CA, USA). Tryptic peptides, obtained by in-solution digestion, were separated on a 12 cmx75 μ m I.D. C18 RP column and eluted peptides were analyzed with LTQ Velos Orbitrap mass spectrometer (Thermo Electron Corp., San Jose, CA, USA). Protein quantification was performed using ProteoIQ software with spectra count data.
Data source location	Cities of Osijek and Rijeka, Croatia
Data accessibility	Filtered data sets are provided as tables with this article

Value of the data

- MS data provide information about alterations in the proteomes of food borne pathogen bacteria *B. subtilis* and *E. coli*, after treatment with newly synthesized quaternary ammonium salts.
- Quantitative gel-free proteomic investigations by LC-ESI MS/MS were used
- Up- and down-regulated proteins in bacterial samples after treatment with four different compounds were identified.
- Protein expression profiles point to the mechanisms of inhibition of used compounds.

1. Data

This report contains the complete list of proteins identified in control (not treated) samples of Gram-positive bacterium *Bacillus subtilis* and Gram-negative bacterium *Escherichia coli* ([Supplementary 1 and 2](#)).

Proteomic data for each bacteria treated with four different disinfectants based on chloride and bromide salts of pyridinium oxime are also provided in the form of tables containing all up- and down regulated proteins compared to the control samples ([Supplementary 3–18](#)). Data presented in this article is related to the research article [\[1\]](#).

2. Experimental design, materials and methods

2.1. Stepwise extraction of proteins

Bacterial cultures of Gram-positive bacterium *B. subtilis* and Gram-negative bacterium *E. coli* used for antimicrobial testing were clinical isolates, isolated from patients and identified according to API tests (API® bioMérieux, Marcy l'Etoile, France).

2.2. Stepwise extraction of proteins

Bacterial suspensions were centrifuged at 5000 g for 20 min, pellets were re-suspended in extraction buffers and subjected to sequential extraction using the ReadyPrep Extraction kit (BioRad, Hercules, CA, USA) as reported [2]. The procedure consisted of three extraction steps, each giving a separate supernatant (protein extracts 1–3). The protein content in recovered supernatants was determined with the Bicinchoninic Acid (BCA) Protein Assay Kit (Pierce, Rockford, IL, USA), according to the manufacturer's instructions. The amount of protein in the supernatant was adjusted to standard protein loads for an LC-ESI-MS/MS analysis (roughly 20 µg/mL) and subjected to a clean-up via precipitation using a ReadyPrep 2D BioRad kit. All experiments were performed in triplicate.

2.3. Identification of proteins with LC-ESI-MS/MS

The “in solution” tryptic digestion of obtained protein extracts was performed as described elsewhere [2]. Tryptic peptides were separated on a 12 cm × 75 µm I.D. C18 RP column (Column Engineering, Ontario, CA, USA) and eluted using a linear gradient starting with 100% solvent A (0.1 M acetic acid in water) to 70% solvent B (0.1 M acetic acid in acetonitrile) over 60 min (Agilent Technologies, Palo Alto, CA, USA). Eluted peptides were introduced into a LTQ Velos Orbitrap mass spectrometer (Thermo Electron Corp., San Jose, CA, USA) and analyzed as described [2,3]. Peptide spectrum matching was performed against species-specific databases that were downloaded from NCBI (links from www.ncbi.nlm.nih.gov/Taxonomy/taxonomyhome.html) using Mascot v.2.3. (Matrix Science, Ltd.).

Parameters used in the database search were: fixed modification of cysteines (S-carbamidomethyl), variable modification of methionines (oxidation), two missed cleavages allowed with trypsin, 20 ppm mass tolerance for precursor ions, and 0.5 Da for fragment ions. Concatenated databases with both “target” and decoy sequences were utilized to estimate the false discovery rate (FDR). Protein identifications were derived from the peptide matches using ProteoIQ v.2. 3.08 [4]. To provide high confidence on peptide sequence assignment and protein identification, data were filtered following stringent criteria: Mowse score of more than 28 for all charge states, at least two peptides per protein, 1% peptide and 1% protein FDR. Protein label free quantification (LFQ) was performed by spectral counting of identified MS/MS spectra using ProteoIQ software.

Acknowledgements

This work was supported by University of Rijeka, scientific Grant no. 13.11.1.2.03.

Transparency document. Supplementary material

Transparency document associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.dib.2017.09.060>.

Appendix A. Supplementary material

Supplementary data associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.dib.2017.09.060>.

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