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Murine CMV Expressing the High Affinity NKG2D Ligand MULT-1: A Model for the Development of Cytomegalovirus-Based Vaccines

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The development of a vaccine against human cytomegalovirus (CMV) has been a subject of long-term medical interest. The research during recent years identified CMV as an attractive vaccine vector against infectious diseases and tumors. The immune response to CMV persists over a lifetime and its unique feature is the inflationary T cell response to certain viral epitopes. CMV encodes numerous genes involved in immunoevasion, which are non-essential for virus growth *in vitro*. The deletion of those genes results in virus attenuation *in vivo*, which enables us to dramatically manipulate its virulence and the immune response. We have previously shown that the murine CMV (MCMV) expressing RAE-1 γ , one of the cellular ligands for the NKG2D receptor, is highly attenuated *in vivo* but retains the ability to induce a strong CD8⁺ T cell response. Here, we demonstrate that recombinant MCMV expressing high affinity NKG2D ligand murine UL16 binding protein-like transcript (MULT-1) (MULT-1MCMV) inserted in the place of its viral inhibitor is dramatically attenuated *in vivo* in a NK cell-dependent manner, both in immunocompetent adult mice and in immunologically immature newborns. MULT-1MCMV was more attenuated than the recombinant virus expressing RAE-1 γ . Despite the drastic sensitivity to innate immune control, MULT-1MCMV induced an efficient CD8⁺ T cell response to viral and vectored antigens. By using *in vitro* assay, we showed that similar to RAE-1 γ MCMV, MULT-1 expressing virus provided strong priming of CD8⁺ T cells. Moreover, MULT-1MCMV was able to induce anti-viral antibodies, which after passing the transplacental barrier protect offspring of immunized mothers from challenge infection. Altogether, this study further supports the concept that CMV expressing NKG2D ligand possesses excellent characteristics to serve as a vaccine or vaccine vector.

Keywords: murine CMV, cytomegalovirus, vaccine, NKG2D, murine UL16 binding protein-like transcript 1, NK cells, CD8⁺ T cells

INTRODUCTION

Human cytomegalovirus (HCMV) is a member of herpesvirus family with high seroprevalence rate worldwide (1). After acute infection, cytomegaloviruses (CMVs) establish life-long latency from which periodic reactivations can occur. In immunocompetent individuals CMV is usually asymptomatic, whereas infection in immunocompromised or immunologically immature individuals can

cause a severe morbidity (2, 3). In healthy individuals, CMV is controlled by the combined effort of innate and adaptive immunity. While NK cell control is critical during the first days of infection, long-term virus control is maintained predominantly by T cells and antibodies (4–6). Despite several decades of intensive work, no HCMV vaccine is still approved (7).

NKG2D is an activating immune receptor expressed on NK cells, activated and antigen-experienced T cells, and a proportion of NKT and $\gamma\delta$ T cells (8). When expressed on NK cells, NKG2D acts as a strong activating receptor, while its engagement on T cells provides co-stimulation (9, 10). Ligands for NKG2D are structurally similar to MHC class I molecules and normally expressed at very low levels. However, upon cellular stress caused by infection or cell transformation, their expression increases leading to NKG2D engagement and activation of immune cells (11, 12). In humans, NKG2D ligands include highly polymorphic MHC-I-related proteins MICA and MICB and UL16-binding proteins (ULBPs) (13). In mice, ligands for NKG2D receptor belong to the family of retinoic acid inducible early transcripts 1 (RAE-1), histocompatibility 60 (H60), and murine UL16 binding protein-like transcript (MULT-1). Although binding to the same receptor, NKG2D ligands differ in their regulation and affinity for the receptor. It is still not fully understood which consequence has the engagement of different NKG2D ligands on the functional outcome of NK and T cell response.

The best evidence that NKG2D engagement plays an important role in immunosurveillance of CMV is the fact that CMV possesses several immunoevasion genes, which prevent the surface expression of NKG2D ligands (14, 15). In MCMV, the majority of these genes belong to the *m145* family of immunoevasins which includes *m145*, *m152*, and *m155* gene products targeting MULT-1, RAE-1, and H60, respectively (16–18). In addition, viral Fc γ R receptor encoded by the *m138* gene has been shown to downmodulate the expression of H60, MULT-1, and RAE-1e ligands (19, 20).

Murine CMV mutants lacking proteins involved in the regulation of NKG2D ligands are attenuated *in vivo* by NK cells. We exploited this knowledge of NKG2D immunoevasion to develop novel CMV-based vaccine vectors. Recombinant MCMV expressing NKG2D ligand RAE-1 γ , inserted in a place of its viral regulator *m152* is severely attenuated *in vivo*, but nevertheless induces strong antigen-specific CD8⁺ T cell response to CMV and vectored antigens, providing long-term protection against bacterial infection and tumors (21–23). Likewise, HCMV expressing ULBP2 in place of its viral regulator is susceptible to control by NK cells, but preserved the ability to stimulate HCMV-specific T cells (24).

In this work, we constructed new MCMV-based vaccine vector expressing NKG2D ligand MULT-1 in place of its viral regulator *m145*. Based on our previous results on RAE-1 γ MCMV, we hypothesized that MULT-1MCMV would also be efficiently controlled while retaining ability to induce potent CD8⁺ T cell response. Indeed, MULT-1MCMV was dramatically attenuated *in vivo* by NK cells and virus was cleared more rapidly than RAE-1 γ MCMV. Nevertheless, MULT-1MCMV induced a strong CD8⁺ T cell response and anti-viral antibodies. This study further supports our previous results showing that recombinant CMVs

expressing NKG2D ligands can be utilized as efficient vaccines and vaccine vectors.

MATERIALS AND METHODS

Construction of Recombinant MCMV Viruses

Wild-type (WT) MCMV refers to a bacterial artificial chromosome (BAC)-derived mouse cytomegalovirus, MW97.01, previously shown to be biologically equivalent to the MCMV Smith strain (VR-1399). Construction of WT MCMV, $\Delta m152$ MCMV, and RAE-1 γ MCMV expressing SIINFEKL was described previously (21, 22, 25). MULT-1MCMV, MULT-1MCMV expressing SIINFEKL and $\Delta m145$ MCMV expressing SIINFEKL were constructed according to published procedure (26). Briefly, for construction of MULT-1MCMV an ORF encoding FLAG-tagged MULT-1 was first cloned into a plasmid containing kanamycin resistance gene (KanR), I-SceI restriction site, and HCMV immediate early promoter (hMIEP) upstream of the cloning site (kind gift from Martin Messerle). The MULT-1 expression cassette containing KanR was PCR amplified using primers 5'-GGGTAAA ACCGCACACAGATGTAGGGCAGACTCTGAGGACCGGT TTTCAACTCCGCGGTTGACATTGATTATTGACT-3' and 5'-GTGAGGGGATTATGCTCTGTTTATTGTC-TCACGACA GACATACAGAGATTTCGGACAGTCATCATGGGATCCCG TCGATGT-3', which contained 60 nucleotides at their 5' ends homologous to the intended integration site in the BAC-cloned MCMV genome, thereby replacing the *m145* ORF following the homologous recombination. To swap the sequence of D^d restricted antigenic *m164*_{167–175} peptide AGPPRYSRI with K^b restricted peptide SIINFEKL, linear DNA fragment was generated using KanR as a template and primers 5'-GCCGTTCCGAAAGG ACTACTGTCTGGACGTGGGGCGCTGACAGTATAATCA ACTTTGAAAACTGAGGATGACGACGATAAGT-3' and 5'-AAGGTCTCCTCGCCCGCTGCCACGATGG-CCTGGTTG TTGACGGCCCAGAACAGTTTTTCAAAGTTGATTATACT GTCAGCGCCCCACCAACCAATTAACCAATTC-3' for PCR amplification. $\Delta m145$ MCMV expressing SIINFEKL was constructed by deletion of MULT-1 from MULT-1MCMV expressing SIINFEKL according to the same procedure using 5'-GGGTAAA ACCGCACACAGATGTAGG-GGCAGACTCTGAGGACCG GTGTTTCAACTCCGCGAGGATGACGACGATAAGTAG-3' and 5'-GATTATGCTCTGTTTATTGTCTCACGACAGACATA CAGAGATTTCGGACAGCGCGGAG-TTGAACACCGG TCCTCAGAGTCTGCCCTACACAACCAATTAACCAA TTCTG-3' primers resulting in *m145* deletion.

Cells and Virus Propagation

BALB/c mouse embryonic fibroblasts (MEF) were grown according to published procedure (27). MEF and SVEC4-10 cells were infected with 1.5 or 3 PFU/cell, respectively. Viruses were propagated on MEF and concentrated by sucrose gradient ultracentrifugation (28). To assess virus replication *in vitro* by multi-step growth kinetics assay, MEF were infected with 0.1 PFU/cell of WT MCMV, RAE-1 γ MCMV, and MULT-1MCMV. Supernatants were harvested at indicated times after infection and virus titers were determined by plaque assay (28).

Mice and Infection

C57BL/6, congenic C57BL/6 (Ly5.1/CD45.1⁺), NKG2D^{-/-} (29), BALB/c, TCR transgenic mice specific for M38 (Maxi) (30), and SIINFEKL (OT-1) (31) were bred under specific pathogen-free conditions at the Faculty of Medicine, University of Rijeka. All experiments performed in this study were approved by the Animal Welfare Committee of the University of Rijeka. Unless otherwise noted, gender matched mice at age of 8–16 weeks were infected with 2×10^5 PFU of tissue culture derived recombinant MCMV either in the footpad (f.p.) or intravenously (i.v.). Newborn BALB/c mice were infected intraperitoneally (i.p.) with 500 PFU of indicated viruses 6 h after birth. Newborn C57BL/6 mice were infected i.p. with 200 or 500 PFU of indicated viruses 24 h post-partum. *In vivo* blocking of NKG2D and depletion of NK cells was performed by i.p. injection of mouse α -mouse NKG2D blocking antibody (generated by Center for Proteomics, University of Rijeka, Faculty of Medicine, clone NKG2D.03) or mouse α -mouse NK1.1 (clone PK136) (32) and rabbit α -asialo GM1 antiserum (α AGM1) (Wako Chemicals), respectively. Viral titers from organs were determined by a plaque assay (28).

Adoptive Transfer

For adoptive transfer experiments C57BL/6 or C57BL/6 CD45.1⁺ mice were immunized f.p. with 2×10^5 PFU of indicated viruses. After 6 weeks, total CD8⁺ T cells were enriched from splenocytes using CD8a⁺ T Cell Isolation Kit (Miltenyi) and sorted on BD FACSAriaII. Adult C57BL/6 recipients were administered i.p. with 250 μ g of depleting NK1.1 antibody (PK136) 1 day prior to sublethal irradiation with 7 Gy. Next day, mice were i.v. injected with 10^6 sorted CD8⁺ T cells. Newborn C57BL/6 mice were i.p. injected with 10^5 sorted CD8⁺ T cells 1 day prior to infection.

In Vitro Stimulation of CD8⁺ T Cells

Bone marrow cells isolated from both femurs and tibias of C57BL/6 mice were differentiated into bone marrow-derived dendritic cells (BMDCs) in the presence of J558 supernatant for 6 days (33). BMDCs were infected with 2 PFU/cell of indicated viruses. After 24 h of infection corresponding amount of either Maxi or OT-1 splenocytes was added at different T:E (BMDCs:CD8⁺) ratios, together with Brefeldin A (Sigma). After 6 h of co-incubation, intracellular staining for IFN- γ production was performed.

Flow Cytometry and Immune Assays

For staining of cell surface expression of NKG2D ligands, mouse NKG2D protein fused with human Fc fragment was used, followed by conjugated donkey α -human IgG Fc secondary antibody (Jackson ImmunoResearch). Alternatively, mouse α -mouse RAE-1 γ (generated by Center for Proteomics, University of Rijeka, Faculty of Medicine, clone RAE-1 γ 0.01) or rat α -mouse MULT-1 (clone 1D6) (18) was used, followed by fluorochrome conjugated goat α -mouse IgG (BD Pharmingen) and goat α -rat IgG F(ab')₂ (Santa Cruz) secondary antibody, respectively. As isotype control irrelevant protein fused with human Fc was used (human PVR-Fc) or antibodies of same isotype originating from the same host, respectively. Splenocytes from immunized C57BL/6 mice were either immediately stained with fluorescently labeled

antibodies for assessment of CD8⁺ T cell phenotype or incubated in the presence of 5 μ g/ml H-2^b restricted custom synthesized peptides of M45 (₉₈₅HGIRNASFI₉₉₃), M38 (₃₁₆SSPPMFRV₃₂₃), and ovalbumin (₂₅₇SIINFEKL₂₆₄) (JPT Peptide Technologies) in presence of Brefeldin A (Sigma) for cytokine production. After 4 h of stimulation, cells were stained for viability, expression of surface markers, and intracellular cytokines. Reagents used in flow cytometry analysis were purchased from eBioscience/Thermo Fischer Scientific and included: Fixable Viability Dye, α CD8 (clone 53-6.7), α CD44 (clone IM7), α CD62L (clone MEL-14), α CD127 (clone SB/199), α IFN γ (clone XMG1.2), and α TNF (clone MP6-XT22) antibodies. Following reagents were obtained through NIH Tetramer Core Facility: H-2K(b) MCMV M38 (₃₁₆SSPPMFRV₃₂₃), H-2L(d) MCMV IE1 (₁₆₈YPHFMPTNL₁₇₆), and H-2D(d) MCMV m164 (₂₅₇AGPPRYSRI₂₆₅). SIINFEKL-specific multimer H-2Kb/SIINFEKL MHC multimer was kindly provided by Dirk Busch (Munich). Samples were analyzed on BD FACSAriaII using FACSDiva and FlowJo (Tree Star, Inc.).

ELISA

For detection of MCMV-specific total IgG in immune sera, high binding ELISA plates were coated with lysate of Δ m138MCMV infected MEF as previously described (34). Shortly, sera of mice were incubated overnight at +4°C, followed by extensive washing and detection with peroxidase conjugated goat α -mouse IgG (H + L) (Jackson ImmunoResearch).

Statistical Analysis

Statistical significance was calculated by unpaired two-tailed Student's *t*-test or Mann-Whitney *U* test for statistical analyses of the virus titers. Differences in MCMV-specific antibody titers in sera of immune animals were analyzed using two-way-ANOVA and Bonferroni *post hoc* test. *P* values less than 0.05 were considered significant. Only statistically significant differences are indicated in figures. All data were analyzed using GraphPad Prism 5 software.

RESULTS

Construction and *In Vitro* Characterization of Recombinant MCMV Expressing MULT-1

MULT-1MCMV was constructed by replacing the *m145* gene, encoding a viral inhibitor of MULT-1, with the gene encoding MULT-1 under the control of hMIEP (Figure 1A). The construction of RAE-1 γ MCMV was described previously (21, 22). To study CD8⁺ T cell response induced by these recombinant viruses against well characterized CD8⁺ T cell epitope, all viruses used in this study expressed the immunodominant K^b epitope SIINFEKL in place of D^d restricted antigenic m164 epitope (Figure 1A). MCMV expressing only SIINFEKL was used as a control (hereby referred as WT MCMV) (25). MULT-1MCMV replication *in vitro* was comparable to WT MCMV and RAE-1 γ MCMV (Figure 1B). To measure the expression of NKG2D ligands on the surface of infected cells, MEF were infected with WT MCMV, Δ m152MCMV, RAE-1 γ MCMV, Δ m145MCMV,

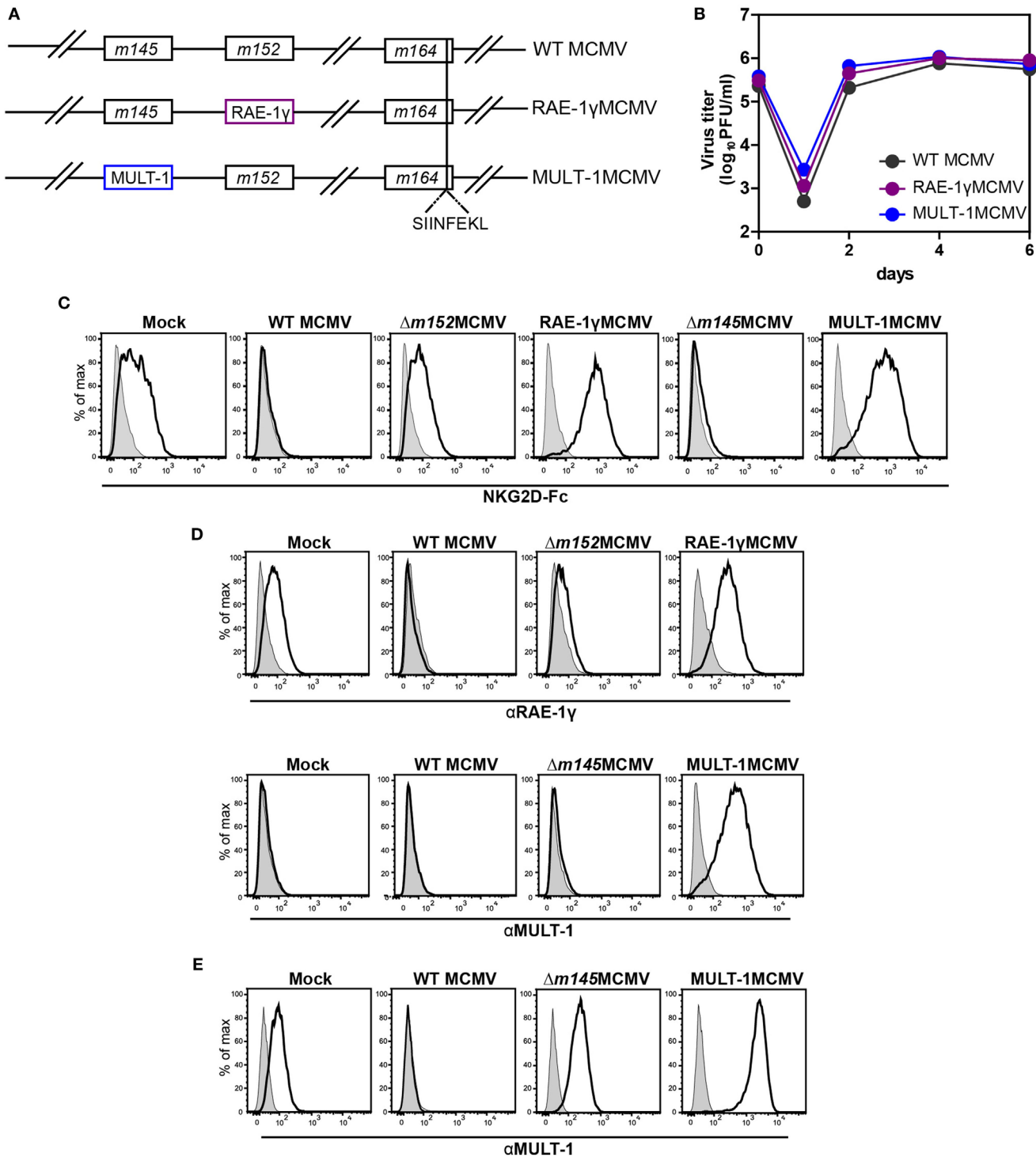


FIGURE 1 | Recombinant viruses used in this study. **(A)** Recombinant murine CMV (MCMV) was made by insertion of genes for NKG2D ligands, RAE-1 γ , and murine UL16 binding protein-like transcript-1 (MULT-1) in place of *m152* and *m145*, respectively. OVA-derived K^b restricted CD8⁺ T cell epitope SIINFEKL was swapped with D^d restricted viral CD8⁺ T cell epitope of *m164* 167AGPPRYSR175. **(B)** Multi-step growth kinetics assay on mouse embryonic fibroblasts (MEF) comparing wild-type (WT) MCMV, RAE-1 γ MCMV, and MULT-1MCMV is shown. **(C,D)** MEFs were infected with 1,5 PFU/cell of indicated viruses and expression of NKG2D ligands was evaluated 24 h after infection by staining either with **(C)** mouse NKG2D-Fc fusion protein (black line) or **(D)** α RAE-1 γ (upper row, black line), α MULT-1 (lower row, black line), and appropriate isotype controls (gray). **(E)** SVEC4-10 cells were infected with 3 PFU/cell of WT MCMV, Δ *m145*MCMV, and MULT-1MCMV for 16 h. Surface expression of MULT-1 was detected with α MULT-1 (black line) or isotype control (gray).

and MULT-1MCMV for 24 h. To assess the expression of exogenous NKG2D ligands as well as endogenous NKG2D ligands affected by the deletion of viral evasins, staining was performed using mouse NKG2D-Fc fusion protein (**Figure 1C**). Expression of inserted NKG2D ligands on the surface of RAE-1 γ MCMV or MULT-1MCMV infected cells was confirmed by staining with specific antibodies against RAE-1 γ and MULT-1, respectively (**Figure 1D**). As previously published, WT MCMV downregulated NKG2D ligands from the cell surface and deletion of *m152* substantially restored RAE-1 surface expression (35). However, infection of MEF with MULT-1MCMV resulted in high surface expression of MULT-1. Furthermore, both RAE-1 γ MCMV and MULT-1MCMV infected cells showed similar binding of NKG2D-Fc (**Figure 1C**). Since MEF constitutively express very low level of MULT-1, the effect of *m145* deletion on the expression of this ligand was hardly detectable. However, the impact of *m145* on the surface expression of MULT-1 was detectable after infecting SVEC4-10 cells that constitutively express MULT-1 (**Figure 1E**) (18, 19). Altogether, these data demonstrate that cells infected with MULT-1MCMV express high levels of MULT-1 on their surface and its expression was not abrogated by function of other viral regulator of MULT-1.

MULT-1MCMV Is Strongly Attenuated in Adult and Neonatal Mice

To determine how expression of high affinity NKG2D ligand MULT-1 affects viral control *in vivo*, BALB/c mice were infected with WT MCMV, RAE-1 γ MCMV, and MULT-1MCMV *i.v.* and viral titers were analyzed at different times after infection. By the day 4 after infection both RAE-1 γ MCMV and MULT-1MCMV were heavily attenuated in spleen, lungs, and liver (**Figures 2A,C**). Moreover, in liver MULT-1MCMV was even more attenuated as compared to RAE-1 γ MCMV (**Figure 2C**). Both RAE-1 γ MCMV and MULT-1MCMV were also attenuated in lungs at day 14 post infection compared to WT MCMV and were completely undetectable in salivary glands (**Figure 2A**). As shown previously (18), $\Delta m145$ MCMV was attenuated as compared to WT MCMV, but attenuation of MULT-1MCMV was even stronger (Figure S1A in Supplementary Material).

We have previously shown that RAE-1 γ MCMV was attenuated in immunologically immature newborns (21). Here, we show that MULT-1MCMV was severely attenuated in newborn mice as well (**Figure 2B**). Surprisingly, MULT-1MCMV was controlled more rapidly than RAE-1 γ MCMV in infected newborns, with virus undetectable at the day 8 after infection.

To identify the immune mechanism responsible for the efficient control of MULT-1MCMV, groups of infected BALB/c mice were treated either with α AGM1 to deplete NK cells or NKG2D blocking antibody. Blocking of NKG2D receptor abolished the differences between mutant viruses and WT MCMV suggesting that early attenuation of recombinant viruses expressing NKG2D ligands is exclusively NKG2D-dependent. Depletion of NK cells by α AGM1 also resulted in the abrogation of differences in viral titer between the groups (**Figure 2C**). It has been well established that NK cells in MCMV resistant C57BL/6 mice control

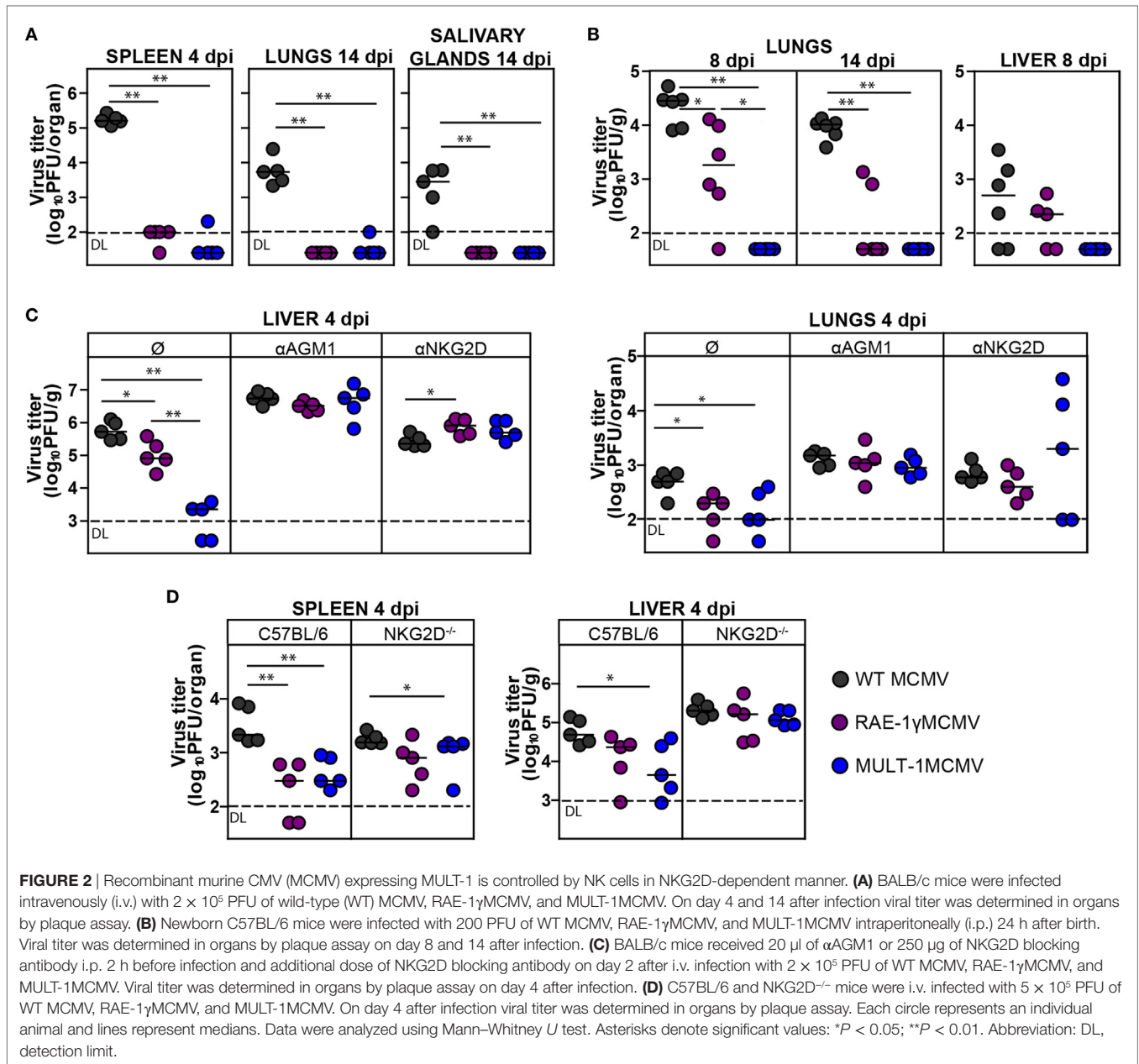
virus *via* a direct recognition of viral *m157* protein by Ly49H activating receptor (36). Notably, viruses expressing NKG2D ligands were more sensitive to NK cell-dependent control even in C57BL/6 mice but this phenotype was lost in NKG2D^{-/-} mice (**Figure 2D**).

MULT-1MCMV Induces a Strong and Functional Antigen-Specific CD8⁺ T Cell Response

We have already shown that MCMV expressing NKG2D ligand RAE-1 γ induces a strong CD8⁺ T cell response despite its attenuation *in vivo* (21, 22). To examine CD8⁺ T cell response after MULT-1MCMV infection, C57BL/6 mice were infected into footpad with WT MCMV, RAE-1 γ MCMV, and MULT-1MCMV. Antigen-specific CD8⁺ T cell response was evaluated in the early (7 days) and late (2 months) phase of infection by measuring IFN- γ and TNF- α production after stimulation with non-inflamatory peptide M45, inflamatory peptide M38, as well as OVA-derived peptide SIINFEKL (**Figure 3A**; Figure S2 in Supplementary Material). At both time points after infection the frequencies and absolute numbers of virus-specific CD8⁺ T cells were comparable or higher in MULT-1MCMV immunized animals compared to WT MCMV. While virus-specific CD8⁺ T cell response was similar in RAE-1 γ MCMV and MULT-1MCMV, RAE-1 γ MCMV induced more CD8⁺ T cells specific for the vectored epitope SIINFEKL. Furthermore, RAE-1 γ MCMV induced more effector memory SIINFEKL-specific CD8⁺ T cells than MULT-1MCMV, while frequency of M38-specific memory subsets was comparable between viruses (Figure S3 in Supplementary Material). Of note, although MULT-1MCMV was attenuated compared to $\Delta m145$ MCMV, both viruses induced similar CD8⁺ T cell response (Figure S1B in Supplementary Material).

To circumvent the impact of a different antigenic load, we performed an *in vitro* stimulation of CD8⁺ T cells. BMDCs were infected with WT MCMV, RAE-1 γ MCMV, and MULT-1MCMV and co-incubated with splenocytes isolated from naive TCR transgenic mice possessing CD8⁺ T cells specific for the M38 epitope (Maxi) or SIINFEKL (OT-1) (**Figure 3B**). MULT-1MCMV infected BMDCs stimulated a higher proportion of Maxi CD8⁺ T cells to produce IFN- γ compared to WT MCMV and equal to RAE-1 γ MCMV. In accordance with data presented in **Figure 3A**, RAE-1 γ MCMV infected BMDCs provided superior stimulation of OT-1 CD8⁺ T cells. However, MULT-1MCMV infected BMDCs stimulated OT-1 CD8⁺ T cells better than WT MCMV. Altogether, MULT-1MCMV induced a robust antigen-specific CD8⁺ T cells response to both viral and vectored epitopes in both acute and latent phase of infection.

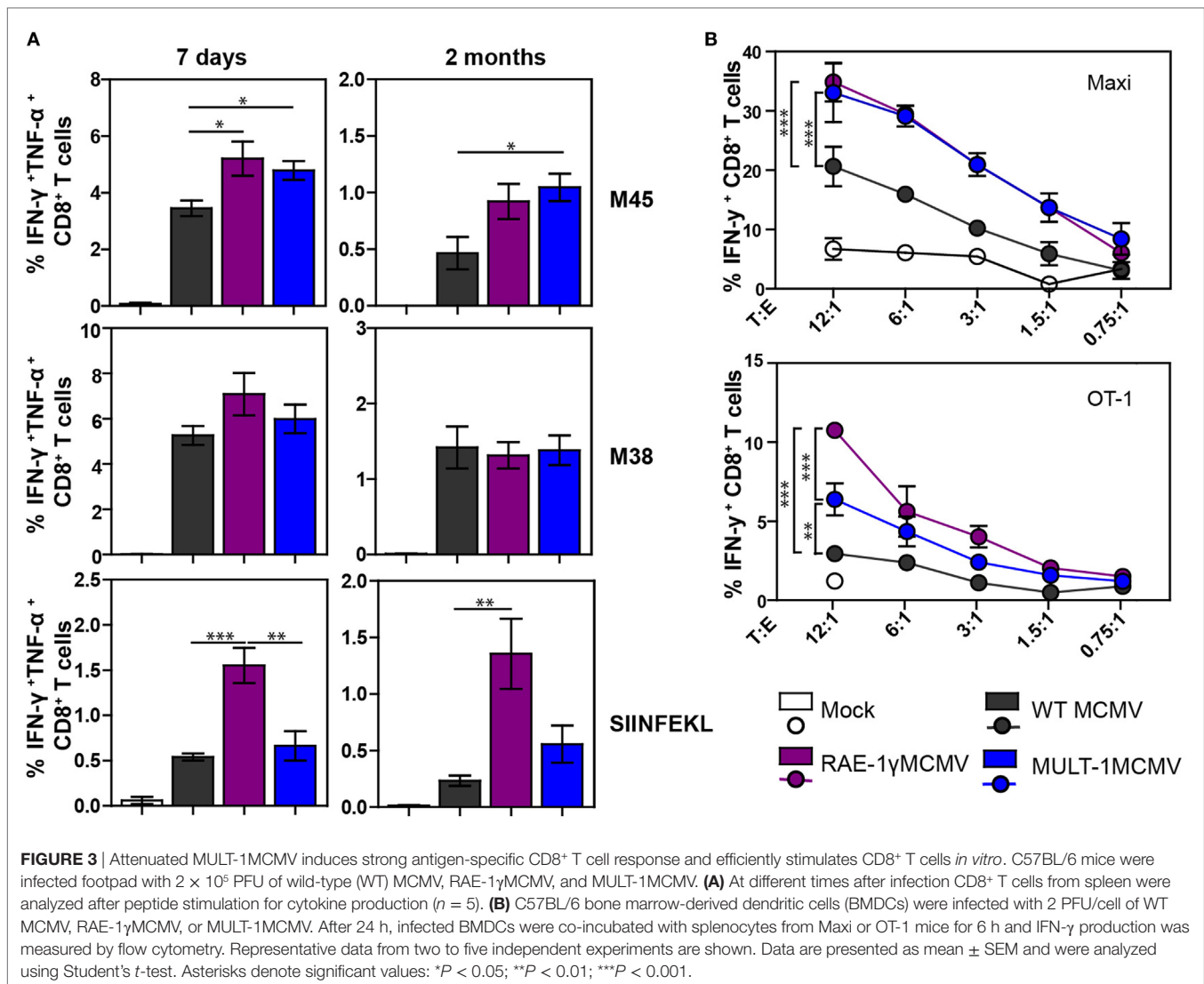
Long-term CD8⁺ T cell memory formation is critical for protection upon challenge infection later in life. To assess the protective capacity of virus-specific CD8⁺ T cells induced by MULT-1MCMV, adoptive transfer experiments were performed. CD8⁺ T cells were sorted from C57BL/6 CD45.1⁺ donor mice latently infected with WT MCMV, RAE-1 γ MCMV, and MULT-1MCMV and adoptively transferred into NK depleted and irradiated C57BL/6 CD45.2⁺ recipients infected



with WT MCMV (Figure 4A). CD8⁺ T cells from MULT-1MCMV immunized donors controlled the infection in lungs and liver of challenged recipients more efficiently than non-immune CD8⁺ T cells and equally efficient as CD8⁺ T cells obtained from WT MCMV and RAE-1 γ MCMV immunization. By using a similar approach, we evaluated the protective capacity of MULT-1MCMV immunization by transferring immune CD8⁺ T cells into newborn mice infected with WT MCMV (Figure 4B). CD8⁺ T cells derived from WT MCMV, RAE-1 γ MCMV, and MULT-1MCMV infected mice provided a similar level of protection in this model as well. Altogether, our results demonstrated that antigen-specific CD8⁺ T cells induced by highly attenuated MULT-1MCMV are functional and protective against MCMV infection.

Immunization of Female Mice With MULT-1MCMV Provides Antibody Mediated Protection to Their Offspring

Induction of protective antibody response is a favorable feature of any vaccine. To determine whether MULT-1MCMV can induce MCMV-specific antibodies in immunized mothers which could protect their offspring from MCMV disease, BALB/c females were infected with WT MCMV, RAE-1 γ MCMV, and MULT-1MCMV 2 weeks prior mating (Figure 5A). Analysis of immune sera showed that MULT-1MCMV immunized mothers, as well as their offspring, had a substantial level of MCMV-specific antibodies (Figure 5B). There was no difference in levels of anti-viral antibodies between MULT-1MCMV compared to WT MCMV.

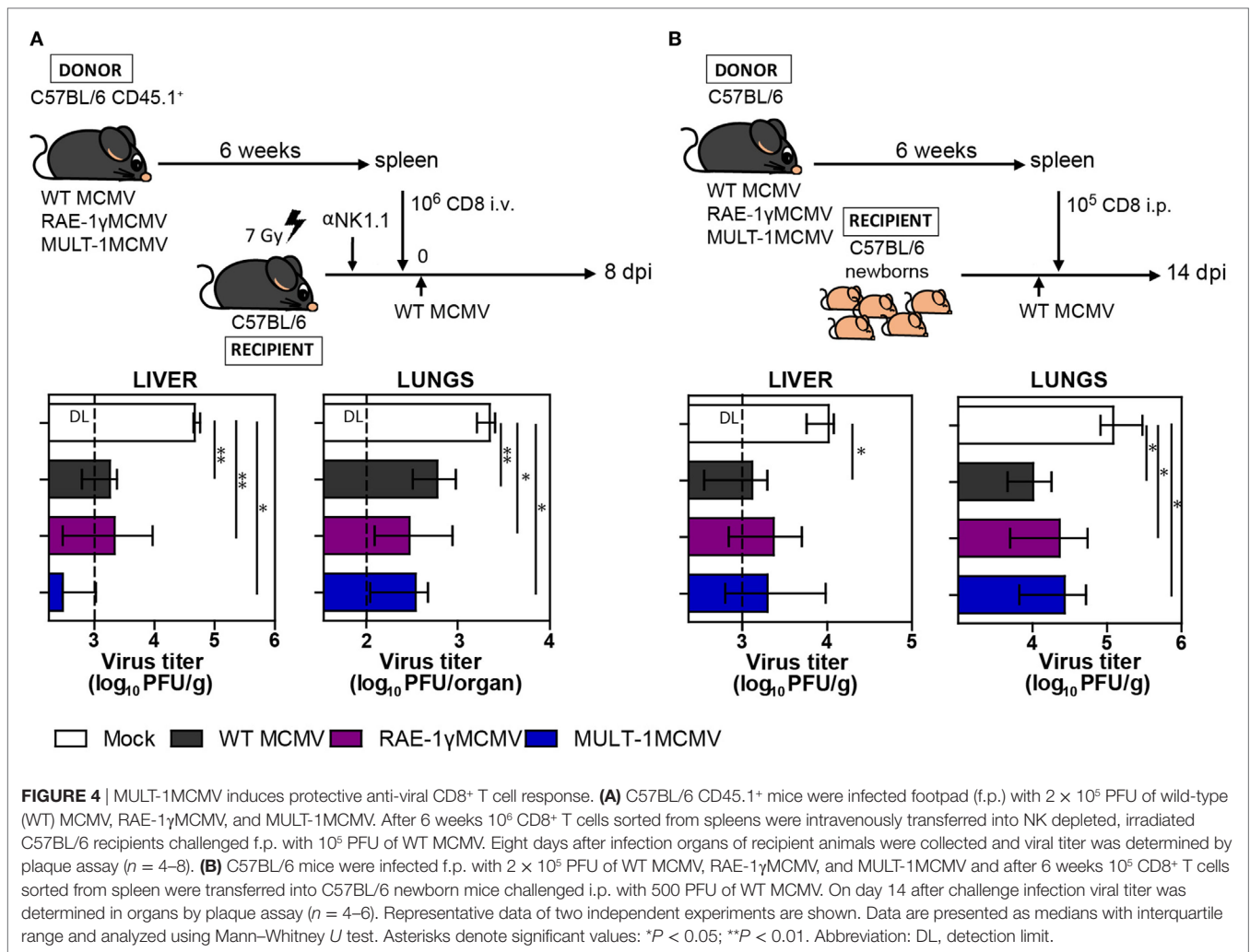


As shown previously, slightly lower antibody titer was observed in mice immunized with RAE-1 γ MCMV (21). Moreover, newborns from MULT-1MCMV immunized mothers were completely protected upon infection with WT MCMV, as well as newborns from WT MCMV and RAE-1 γ MCMV immunized dams (Figure 5C). Altogether, these data show that despite a strong attenuation, MULT-1MCMV immunization of mothers induces a strong production of anti-viral antibodies, which can pass the placenta and protect their offspring from MCMV disease.

DISCUSSION

After resolution of primary CMV infection, CD8⁺ T cells specific for certain immunodominant epitopes are not maintained as a low abundant memory population, but rather gradually increase in frequency acquiring an effector-like phenotype, a phenomenon known as “memory inflation” (37, 38). This characteristic of CMV induced CD8⁺ T cell response could be exploited in the development of CD8⁺ T cell-based live vaccines

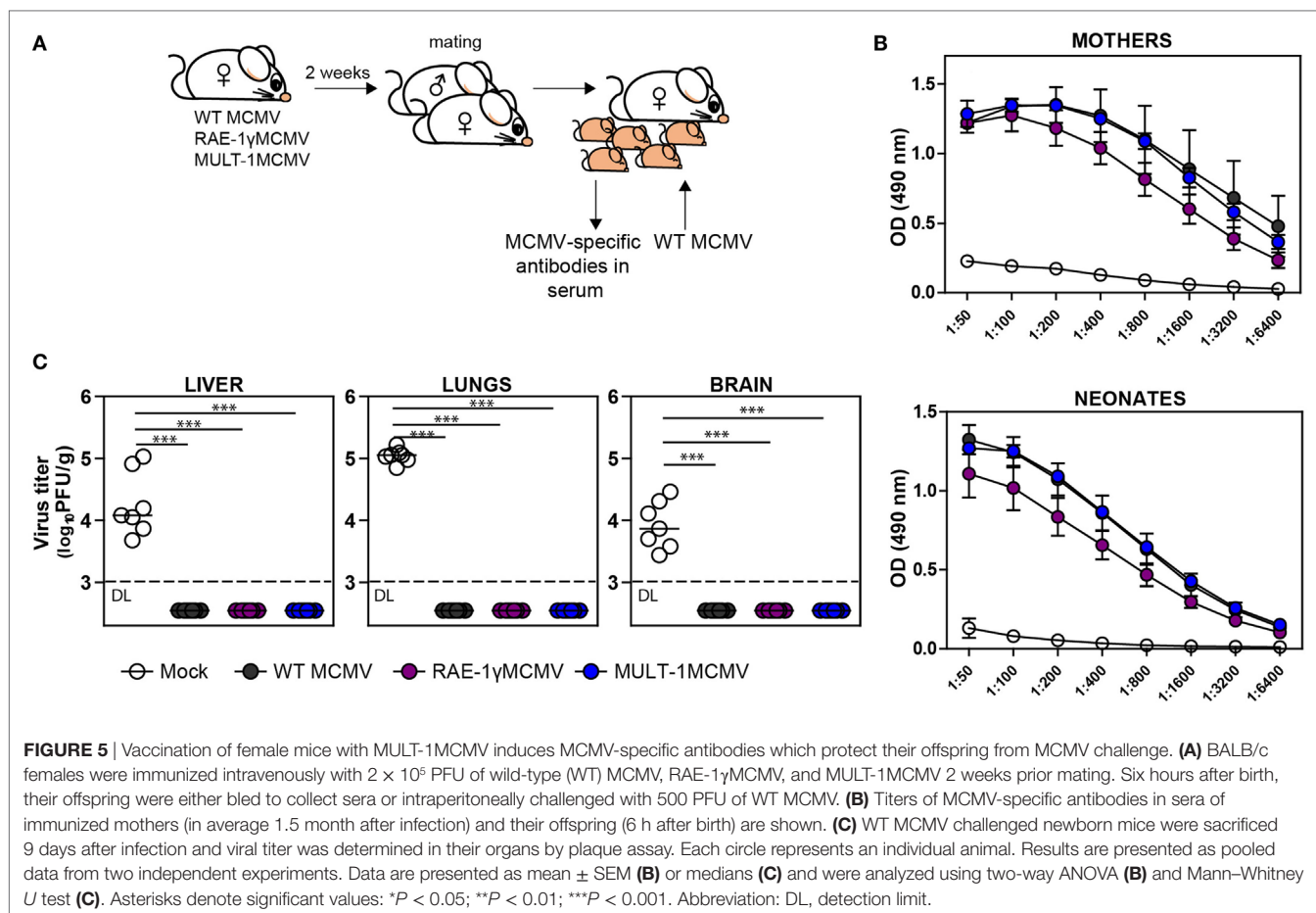
(39–42). Live attenuated vaccines imitate natural infection by inducing a broad cellular and humoral response to variety of antigens, which makes them superior to subunit vaccines. Still, there are concerns about using live virus as a vaccine as it might cause unwanted virulence, especially in high-risk individuals such as immunocompromised patients. CMV has many non-essential genes involved in subversion of immune response and the deletion of those genes enables the manipulation of its virulence and the quality of immune response. In our previous studies we took advantage of virus lacking the NKG2D immunoevasin to generate recombinant immunologically attenuated vaccine vector RAE-1 γ MCMV that retained the ability to induce a strong adaptive response (21–23). NKG2D ligands are differently expressed and regulated in tissues, they differ in the affinity for the receptor and certain ligands are restricted to the particular mouse strains (11, 43–45). MULT-1 has the highest affinity for the receptor among all mouse NKG2D ligands and its transcripts were found in most of the healthy tissues, but its protein expression was strictly regulated on posttranslational



level (46). To exploit those differences in the context of a MCMV-based vaccine, we constructed MULT-1MCMV and compared it with WT MCMV and RAE-1 γ MCMV. MULT-1MCMV was highly immunologically attenuated, but nevertheless induced a strong and protective antigen-specific CD8⁺ T cell and antibody response. Thus, this study further supports the idea that attenuated recombinant CMVs expressing NKG2D ligands can be used as efficient vaccines and vaccine vectors.

It is well known that NK cell response can also shape the adaptive immune response (47). The most extensively studied example of strong NK control of MCMV infection is the direct recognition of viral protein m157 by the activating NK receptor Ly49H, leading to efficient virus control (36, 48). In most cases, a strong NK control would impair CD8⁺ T cell responses presumably due to the lower viral burden and cytokine milieu diminishing antigen presentation (49, 50). Indeed, during MCMV infection the lower viral burden reduces the magnitude of long-term CD8⁺ T cell response but does not impact its kinetics (51, 52). Paradoxically, it has been shown that blocking viral replication increases CD8⁺ T cell response to non-inflamatory epitopes presented by preserved dendritic cells (DCs) which would otherwise be depleted due to type I IFNs (53, 54). Interestingly, this effect was

not evident in the population of inflamatory CD8⁺ T cells. While conventional non-inflamatory CD8⁺ T cells are preferentially primed by cross-presented antigens on DCs (55), inflamatory CD8⁺ T cells are originating from cells primed early in infection as well as from constantly replenished short-lived effectors (56) maintained at high levels by sporadic virus reactivations in non-hematopoietic cells (30, 57). Differential requirements in generation of non-inflamatory and inflamatory CD8⁺ T cells in MCMV infection might explain these findings (58). Here we show that despite efficient NK-mediated virus control, MULT-1MCMV induces CD8⁺ T cell response of similar or greater magnitude compared to WT MCMV to both non-inflamatory and inflamatory epitopes. While all viruses induced comparable CD8⁺ T cell response to viral antigens, RAE-1 γ MCMV induced a higher number of SIINFEKL-specific CD8⁺ T cells. Somewhat better response to non-inflamatory epitope M45 in mice infected with recombinant viruses expressing NKG2D ligands could be explained either by a better cross-presentation due to preserved DCs in absence of strong infection or immune function of these proteins (21, 22, 54). The strong engagement of NKG2D and activation of NK cells might result in an environment favoring antigen presentation. Alternatively, the co-stimulation signal



by engagement of NKG2D on CD8⁺ T cells could rescue T cell responses which would otherwise be weaker due to strong viral control and action of other viral evasins. The importance of CD8⁺ T cell co-stimulation is evident from the fact that TCR signaling without an appropriate co-stimulation signal drives CD8⁺ T cells to the state of anergy rather than activation (59). CMVs have developed multiple mechanisms to evade CD8⁺ T cell recognition of infected cells. Indeed, MCMV possess several genes encoding proteins which interfere with antigen presentation by downregulating the expression of MHC I molecules (6). Ligands for the major T cell co-stimulatory receptor CD28 are also targeted by viral immunoevasion (59–61). CD28/B7-mediated co-stimulation is indispensable in acute infection and for the establishment of CD8⁺ T cell memory in MCMV (62). In LCMV infection, the absence of B7-mediated co-stimulation can be substituted with other co-stimulatory pathways (63). Though this was not shown for CD8⁺ T cell response in herpes viruses, it suggests that under certain conditions co-stimulatory pathways might act compensatory to each other. Our *in vitro* studies showed that RAE-1 γ MCMV and MULT-1MCMV infected BMDCs have an improved capacity to stimulate CD8⁺ T cells compared to WT MCMV. The engagement of co-stimulatory receptor NKG2D on CD8⁺ T cells was reported to promote proliferation and cytotoxic capacity of antigen-specific CD8⁺ T cells in various experimental settings (64–66), as well as rescue memory of unhelped CD8⁺ T cells (67). Moreover, it was shown

recently that engagement of NKG2D on NK cells augments their expansion during MCMV infection (68). Therefore, we hypothesize that co-stimulatory function of NKG2D on CD8⁺ T cells might overcome multiple ways of viral interference with antigen presentation and downregulation of co-stimulatory molecules resulting in a better stimulation and improved CD8⁺ T cell responses *in vivo*.

Apart from prompting durable CD8⁺ T cell memory, the ability to induce humoral response is another vital feature of CMV vaccine, since antigen-specific antibodies are able to cross the placental barrier and are the first to protect the fetus and newborns against congenital infection (69, 70). Despite the lower antigenic load, MULT-1MCMV immunization induced MCMV-specific antibodies capable of protecting offspring against a challenge infection.

Another interesting finding of this study was the rapid attenuation of MULT-1MCMV pronounced in perinatally infected newborns. This is in agreement with our recent study showing that MULT-1MCMV failed to reach the brain of mice infected as newborns resulting in the absence of brain inflammation and establishment of tissue-resident memory CD8⁺ T cells (71). NK cells are immature in newborns, at least partially due to the presence of high levels of immune suppressive TGF- β , which makes newborns highly susceptible to viral infections (72). Since NKG2D is expressed early in ontogeny of NK cells (73), we speculate that the engagement of NKG2D with a high

affinity ligand might overcome the suppressive environment in newborn mice, but the exact mechanism of MULT-1/MCMV control in newborns remains elusive. Yet, this study demonstrates the feasibility of CMV-based vaccine vectors to be used even in newborns.

Altogether, we show for the first time that MCMV expressing the high affinity NKG2D ligand MULT-1 has numerous favorable features of a vaccine such as being highly attenuated, but still able to induce both cellular and humoral adaptive immunity. We have established the vaccine properties of MULT-1/MCMV against MCMV infection, which does not exclude the prospect to test its capacity as a vector vaccine. In conclusion, by comparing recombinant viruses expressing different NKG2D ligands, we confirmed the dual role of NKG2D in mediating a strong virus control, while retaining CD8⁺ T cell response equivalent or better than WT virus infection, which can serve as a model approach for the development of a similar HCMV-based vaccine vector.

ETHICS STATEMENT

This study was carried out in accordance with the recommendations of Regulations on the protection of animals used for scientific purposes (Official Gazette of the Republic of Croatia, 55/2013). Ethics Committee of the Veterinary Department of the Ministry of Agriculture and Animal Welfare Committee of the University of Rijeka Faculty of Medicine approved all animal experiments.

REFERENCES

- Boppana SB, Fowler KB. Chapter 36: Persistence in the population: epidemiology and transmission. In: Arvin A, Campadelli-Fiume G, Mocarski E, Moore PS, Roizman B, Whitley R, Yamanishi K, editors. *Human Herpesviruses: Biology, Therapy, and Immunoprophylaxis*. Cambridge: Cambridge University Press (2007).
- Rubin RH. The indirect effects of cytomegalovirus infection on the outcome of organ transplantation. *JAMA* (1989) 261(24):3607–9. doi:10.1001/jama.1989.03420240121038
- Pass RF, Fowler KB, Boppana SB, Britt WJ, Stagno S. Congenital cytomegalovirus infection following first trimester maternal infection: symptoms at birth and outcome. *J Clin Virol* (2006) 35(2):216–20. doi:10.1016/j.jcv.2005.09.015
- Jonjic S, Pavic I, Polic B, Crnkovic I, Lucin P, Koszinowski UH. Antibodies are not essential for the resolution of primary cytomegalovirus infection but limit dissemination of recurrent virus. *J Exp Med* (1994) 179(5):1713–7. doi:10.1084/jem.179.5.1713
- Polic B, Hengel H, Krmpotic A, Trgovcich J, Pavic I, Luccaroni P, et al. Hierarchical and redundant lymphocyte subset control precludes cytomegalovirus replication during latent infection. *J Exp Med* (1998) 188(6):1047–54. doi:10.1084/jem.188.6.1047
- Reddehase MJ. Antigens and immunoevasins: opponents in cytomegalovirus immune surveillance. *Nat Rev Immunol* (2002) 2(11):831–44. doi:10.1038/nri932
- Schleiss MR. Cytomegalovirus vaccines under clinical development. *J Virus Erad* (2016) 2(4):198–207.
- Raulet DH. Roles of the NKG2D immunoreceptor and its ligands. *Nat Rev Immunol* (2003) 3(10):781–90. doi:10.1038/nri1199
- Diefenbach A, Tomasello E, Lucas M, Jamieson AM, Hsia JK, Vivier E, et al. Selective associations with signaling proteins determine stimulatory versus costimulatory activity of NKG2D. *Nat Immunol* (2002) 3(12):1142–9. doi:10.1038/ni858

AUTHOR CONTRIBUTIONS

LH and IB designed the study, performed the experiments, analyzed the data, and wrote the manuscript. TJ and JR designed and generated recombinant viruses. VL and SJu contributed to performing experiments and critical reading of the manuscript. AK and SJo designed and oversaw the study and wrote the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at <https://www.frontiersin.org/articles/10.3389/fimmu.2018.00991/full#supplementary-material>.

- Gilfillan S, Ho EL, Cella M, Yokoyama WM, Colonna M. NKG2D recruits two distinct adaptors to trigger NK cell activation and costimulation. *Nat Immunol* (2002) 3(12):1150–5. doi:10.1038/ni857
- Cerwenka A, Bakker AB, McClanahan T, Wagner J, Wu J, Phillips JH, et al. Retinoic acid early inducible genes define a ligand family for the activating NKG2D receptor in mice. *Immunity* (2000) 12(6):721–7. doi:10.1016/S1074-7613(00)80222-8
- Gasser S, Orsulic S, Brown EJ, Raulet DH. The DNA damage pathway regulates innate immune system ligands of the NKG2D receptor. *Nature* (2005) 436(7054):1186–90. doi:10.1038/nature03884
- Champsaur M, Lanier LL. Effect of NKG2D ligand expression on host immune responses. *Immunol Rev* (2010) 235(1):267–85. doi:10.1111/j.0105-2896.2010.00893.x
- Sun JC, Lanier LL. The natural selection of herpesviruses and virus-specific NK cell receptors. *Viruses* (2009) 1(3):362. doi:10.3390/v1030362
- Slavuljica I, Krmpotic A, Jonjic S. Manipulation of NKG2D ligands by cytomegaloviruses: impact on innate and adaptive immune response. *Front Immunol* (2011) 2:85. doi:10.3389/fimmu.2011.00085
- Lodoen MB, Abenes G, Umamoto S, Houchins JP, Liu F, Lanier LL. The cytomegalovirus m155 gene product subverts natural killer cell antiviral protection by disruption of H60-NKG2D interactions. *J Exp Med* (2004) 200(8):1075–81. doi:10.1084/jem.20040583
- Hasan M, Krmpotic A, Ruzsics Z, Bubic I, Lenac T, Halenius A, et al. Selective down-regulation of the NKG2D ligand H60 by mouse cytomegalovirus m155 glycoprotein. *J Virol* (2005) 79(5):2920–30. doi:10.1128/JVI.79.5.2920-2930.2005
- Krmpotic A, Hasan M, Loewendorf A, Saulig T, Halenius A, Lenac T, et al. NK cell activation through the NKG2D ligand MULT-1 is selectively prevented by the glycoprotein encoded by mouse cytomegalovirus gene m145. *J Exp Med* (2005) 201(2):211–20. doi:10.1084/jem.20041617
- Lenac T, Budt M, Arapovic J, Hasan M, Zimmermann A, Simic H, et al. The herpesviral Fc receptor fcr-1 down-regulates the NKG2D ligands MULT-1 and H60. *J Exp Med* (2006) 203(8):1843–50. doi:10.1084/jem.20060514

20. Arapovic J, Lenac Rovis T, Reddy AB, Krmpotic A, Jonjic S. Promiscuity of MCMV immunoevasin of NKG2D: m138/fcr-1 down-modulates RAE-1epsilon in addition to MULT-1 and H60. *Mol Immunol* (2009) 47(1):114–22. doi:10.1016/j.molimm.2009.02.010
21. Slavuljica I, Busche A, Babic M, Mitrovic M, Gasparovic I, Cekinovic D, et al. Recombinant mouse cytomegalovirus expressing a ligand for the NKG2D receptor is attenuated and has improved vaccine properties. *J Clin Invest* (2010) 120(12):4532–45. doi:10.1172/JCI43961
22. Trsan T, Busche A, Abram M, Wensveen FM, Lemmermann NA, Arapovic M, et al. Superior induction and maintenance of protective CD8 T cells in mice infected with mouse cytomegalovirus vector expressing RAE-1gamma. *Proc Natl Acad Sci U S A* (2013) 110(41):16550–5. doi:10.1073/pnas.1310215110
23. Trsan T, Vukovic K, Filipovic P, Brizic AL, Lemmermann NAW, Schober K, et al. Cytomegalovirus vector expressing RAE-1gamma induces enhanced anti-tumor capacity of murine CD8(+) T cells. *Eur J Immunol* (2017) 47(8):1354–67. doi:10.1002/eji.201746964
24. Tomic A, Varanasi PR, Golemac M, Malic S, Riese P, Borst EM, et al. Activation of innate and adaptive immunity by a recombinant human cytomegalovirus strain expressing an NKG2D ligand. *PLoS Pathog* (2016) 12(12):e1006015. doi:10.1371/journal.ppat.1006015
25. Lemmermann NA, Gergely K, Bohm V, Deegen P, Daubner T, Reddehase MJ. Immune evasion proteins of murine cytomegalovirus preferentially affect cell surface display of recently generated peptide presentation complexes. *J Virol* (2010) 84(3):1221–36. doi:10.1128/JVI.02087-09
26. Tischer BK, Smith GA, Osterrieder N. En passant mutagenesis: a two step markerless red recombination system. *Methods Mol Biol* (2010) 634:421–30. doi:10.1007/978-1-60761-652-8_30
27. Jonjic S, Krmpotic A, Arapovic J, Koszinowski UH. Dissection of the antiviral NK cell response by MCMV mutants. *Methods Mol Biol* (2008) 415:127–49. doi:10.1007/978-1-59745-570-1_8
28. Brune W, Hengel H, Koszinowski UH. A mouse model for cytomegalovirus infection. *Curr Protoc Immunol* (2001) Chapter 19:Unit19.17. doi:10.1002/0471142735.im1907s43
29. Zafirova B, Mandaric S, Antulov R, Krmpotic A, Jonsson H, Yokoyama WM, et al. Altered NK cell development and enhanced NK cell-mediated resistance to mouse cytomegalovirus in NKG2D-deficient mice. *Immunity* (2009) 31(2):270–82. doi:10.1016/j.immuni.2009.06.017
30. Torti N, Walton SM, Brocker T, Rulicke T, Oxenius A. Non-hematopoietic cells in lymph nodes drive memory CD8 T cell inflation during murine cytomegalovirus infection. *PLoS Pathog* (2011) 7(10):e1002313. doi:10.1371/journal.ppat.1002313
31. Hogquist KA, Jameson SC, Heath WR, Howard JL, Bevan MJ, Carbone FR. T cell receptor antagonist peptides induce positive selection. *Cell* (1994) 76(1):17–27. doi:10.1016/0092-8674(94)90169-4
32. Koo GC, Peppard JR. Establishment of monoclonal anti-Nk-1.1 antibody. *Hybridoma* (1984) 3(3):301–3. doi:10.1089/hyb.1984.3.301
33. Inaba K, Inaba M, Romani N, Aya H, Deguchi M, Ikehara S, et al. Generation of large numbers of dendritic cells from mouse bone marrow cultures supplemented with granulocyte/macrophage colony-stimulating factor. *J Exp Med* (1992) 176(6):1693–702. doi:10.1084/jem.176.6.1693
34. Jonjic S, Pavic I, Lucin P, Rukavina D, Koszinowski UH. Efficacious control of cytomegalovirus infection after long-term depletion of CD8+ T lymphocytes. *J Virol* (1990) 64(11):5457–64.
35. Lodoen M, Ogasawara K, Hamerman JA, Arase H, Houchins JP, Mocarski ES, et al. NKG2D-mediated natural killer cell protection against cytomegalovirus is impaired by viral gp40 modulation of retinoic acid early inducible 1 gene molecules. *J Exp Med* (2003) 197(10):1245–53. doi:10.1084/jem.20021973
36. Arase H, Mocarski ES, Campbell AE, Hill AB, Lanier LL. Direct recognition of cytomegalovirus by activating and inhibitory NK cell receptors. *Science* (2002) 296(5571):1323–6. doi:10.1126/science.1070884
37. Holtappels R, Pahl-Seibert MF, Thomas D, Reddehase MJ. Enrichment of immediate-early 1 (m123/pp89) peptide-specific CD8 T cells in a pulmonary CD62L(lo) memory-effector cell pool during latent murine cytomegalovirus infection of the lungs. *J Virol* (2000) 74(24):11495–503. doi:10.1128/JVI.74.24.11495-11503.2000
38. Klenerman P, Oxenius A. T cell responses to cytomegalovirus. *Nat Rev Immunol* (2016) 16(6):367–77. doi:10.1038/nri.2016.38
39. Karrer U, Wagner M, Siervo S, Oxenius A, Hengel H, Dumrese T, et al. Expansion of protective CD8+ T-cell responses driven by recombinant cytomegaloviruses. *J Virol* (2004) 78(5):2255–64. doi:10.1128/JVI.78.5.2255-2264.2004
40. Hansen SG, Vieville C, Whizin N, Coyne-Johnson L, Siess DC, Drummond DD, et al. Effector memory T cell responses are associated with protection of rhesus monkeys from mucosal simian immunodeficiency virus challenge. *Nat Med* (2009) 15(3):293–9. doi:10.1038/nm.1935
41. Tsuda Y, Caposio P, Parkins CJ, Botto S, Messaoudi I, Cicin-Sain L, et al. A replicating cytomegalovirus-based vaccine encoding a single Ebola virus nucleoprotein CTL epitope confers protection against Ebola virus. *PLoS Negl Trop Dis* (2011) 5(8):e1275. doi:10.1371/journal.pntd.0001275
42. Qiu Z, Huang H, Grenier JM, Perez OA, Smilowitz HM, Adler B, et al. Cytomegalovirus-based vaccine expressing a modified tumor antigen induces potent tumor-specific CD8(+) T-cell response and protects mice from melanoma. *Cancer Immunol Res* (2015) 3(5):536–46. doi:10.1158/2326-6066.CIR-14-0044
43. Nomura M, Zou Z, Joh T, Takihara Y, Matsuda Y, Shimada K. Genomic structures and characterization of Rael family members encoding GPI-anchored cell surface proteins and expressed predominantly in embryonic mouse brain. *J Biochem* (1996) 120(5):987–95. doi:10.1093/oxfordjournals.jbchem.a021517
44. Carayannopoulos LN, Naidenko OV, Fremont DH, Yokoyama WM. Cutting edge: murine UL16-binding protein-like transcript 1: a newly described transcript encoding a high-affinity ligand for murine NKG2D. *J Immunol* (2002) 169(8):4079–83. doi:10.4049/jimmunol.169.8.4079
45. Takada A, Yoshida S, Kajikawa M, Miyatake Y, Tomaru U, Sakai M, et al. Two novel NKG2D ligands of the mouse H60 family with differential expression patterns and binding affinities to NKG2D. *J Immunol* (2008) 180(3):1678–85. doi:10.4049/jimmunol.180.3.1678
46. Nice TJ, Coscoy L, Raulet DH. Posttranslational regulation of the NKG2D ligand Multi1 in response to cell stress. *J Exp Med* (2009) 206(2):287–98. doi:10.1084/jem.20081335
47. Cook KD, Waggoner SN, Whitmire JK. NK cells and their ability to modulate T cells during virus infections. *Crit Rev Immunol* (2014) 34(5):359–88. doi:10.1615/CritRevImmunol.2014010604
48. Smith HR, Heusel JW, Mehta IK, Kim S, Dorner BG, Naidenko OV, et al. Recognition of a virus-encoded ligand by a natural killer cell activation receptor. *Proc Natl Acad Sci U S A* (2002) 99(13):8826–31. doi:10.1073/pnas.092258599
49. Andrews DM, Estcourt MJ, Andoniou CE, Wikstrom ME, Khong A, Voigt V, et al. Innate immunity defines the capacity of antiviral T cells to limit persistent infection. *J Exp Med* (2010) 207(6):1333–43. doi:10.1084/jem.20091193
50. Mitrovic M, Arapovic J, Jordan S, Fodil-Cornu N, Ebert S, Vidal SM, et al. The NK cell response to mouse cytomegalovirus infection affects the level and kinetics of the early CD8(+) T-cell response. *J Virol* (2012) 86(4):2165–75. doi:10.1128/JVI.06042-11
51. Andrews DM, Andoniou CE, Fleming P, Smyth MJ, Degli-Esposti MA. The early kinetics of cytomegalovirus-specific CD8+ T-cell responses are not affected by antigen load or the absence of perforin or gamma interferon. *J Virol* (2008) 82(10):4931–7. doi:10.1128/JVI.02127-07
52. Redeker A, Welten SP, Arens R. Viral inoculum dose impacts memory T-cell inflation. *Eur J Immunol* (2014) 44(4):1046–57. doi:10.1002/eji.201343946
53. Snyder CM, Cho KS, Bonnett EL, Allan JE, Hill AB. Sustained CD8+ T cell memory inflation after infection with a single-cycle cytomegalovirus. *PLoS Pathog* (2011) 7(10):e1002295. doi:10.1371/journal.ppat.1002295
54. Loo CP, Snyder CM, Hill AB. Blocking virus replication during acute murine cytomegalovirus infection paradoxically prolongs antigen presentation and increases the CD8+ T cell response by preventing type I IFN-dependent depletion of dendritic cells. *J Immunol* (2017) 198(1):383–93. doi:10.4049/jimmunol.1600478
55. Snyder CM, Allan JE, Bonnett EL, Doom CM, Hill AB. Cross-presentation of a spread-defective MCMV is sufficient to prime the majority of virus-specific CD8+ T cells. *PLoS One* (2010) 5(3):e9681. doi:10.1371/journal.pone.0009681
56. Snyder CM, Cho KS, Bonnett EL, van Dommelen S, Shellam GR, Hill AB. Memory inflation during chronic viral infection is maintained by continuous production of short-lived, functional T cells. *Immunity* (2008) 29(4):650–9. doi:10.1016/j.immuni.2008.07.017
57. Seckert CK, Schader SI, Ebert S, Thomas D, Freitag K, Renzaho A, et al. Antigen-presenting cells of haematopoietic origin prime cytomegalovirus-specific CD8 T-cells but are not sufficient for driving memory inflation during viral latency. *J Gen Virol* (2011) 92(Pt 9):1994–2005. doi:10.1099/vir.0.031815-0

58. Torti N, Walton SM, Murphy KM, Oxenius A. Batf3 transcription factor-dependent DC subsets in murine CMV infection: differential impact on T-cell priming and memory inflation. *Eur J Immunol* (2011) 41(9):2612–8. doi:10.1002/eji.201041075
59. Chen L, Flies DB. Molecular mechanisms of T cell co-stimulation and co-inhibition. *Nat Rev Immunol* (2013) 13(4):227–42. doi:10.1038/nri3405
60. Loewendorf A, Kruger C, Borst EM, Wagner M, Just U, Messerle M. Identification of a mouse cytomegalovirus gene selectively targeting CD86 expression on antigen-presenting cells. *J Virol* (2004) 78(23):13062–71. doi:10.1128/JVI.78.23.13062-13071.2004
61. Mintern JD, Klemm EJ, Wagner M, Paquet ME, Napier MD, Kim YM, et al. Viral interference with B7-1 costimulation: a new role for murine cytomegalovirus fc receptor-1. *J Immunol* (2006) 177(12):8422–31. doi:10.4049/jimmunol.177.12.8422
62. Arens R, Loewendorf A, Redeker A, Sierro S, Boon L, Klenerman P, et al. Differential B7-CD28 costimulatory requirements for stable and inflationary mouse cytomegalovirus-specific memory CD8 T cell populations. *J Immunol* (2011) 186(7):3874–81. doi:10.4049/jimmunol.1003231
63. Welten SP, Redeker A, Franken KL, Oduro JD, Ossendorp F, Cicin-Sain L, et al. The viral context instructs the redundancy of costimulatory pathways in driving CD8(+) T cell expansion. *Elife* (2015) 4:e07486. doi:10.7554/eLife.07486
64. Groh V, Rhinehart R, Randolph-Habecker J, Topp MS, Riddell SR, Spies T. Costimulation of CD8 α T cells by NKG2D via engagement by MIC induced on virus-infected cells. *Nat Immunol* (2001) 2(3):255–60. doi:10.1038/85321
65. Markiewicz MA, Carayannopoulos LN, Naidenko OV, Matsui K, Burack WR, Wise EL, et al. Costimulation through NKG2D enhances murine CD8+ CTL function: similarities and differences between NKG2D and CD28 costimulation. *J Immunol* (2005) 175(5):2825–33. doi:10.4049/jimmunol.175.5.2825
66. Kavazovic I, Lenart M, Jelenc V, Jurkovic S, Lemmermann NAW, Jonjic S, et al. NKG2D stimulation of CD8(+) T cells during priming promotes their capacity to produce cytokines in response to viral infection in mice. *Eur J Immunol* (2017) 47(7):1123–35. doi:10.1002/eji.201646805
67. Zloza A, Kohlhapp FJ, Lyons GE, Schenkel JM, Moore TV, Lacey AT, et al. NKG2D signaling on CD8(+) T cells represses T-bet and rescues CD4-unhelped CD8(+) T cell memory recall but not effector responses. *Nat Med* (2012) 18(3):422–8. doi:10.1038/nm.2683
68. Nabekura T, Gotthardt D, Niizuma K, Trsan T, Jenus T, Jonjic S, et al. Cutting edge: NKG2D signaling enhances NK cell responses but alone is insufficient to drive expansion during mouse cytomegalovirus infection. *J Immunol* (2017) 199(5):1567–71. doi:10.4049/jimmunol.1700799
69. Boppa SB, Britt WJ. Antiviral antibody responses and intrauterine transmission after primary maternal cytomegalovirus infection. *J Infect Dis* (1995) 171(5):1115–21. doi:10.1093/infdis/171.5.1115
70. Furione M, Rognoni V, Sarasini A, Zavattoni M, Lillier D, Gerna G, et al. Slow increase in IgG avidity correlates with prevention of human cytomegalovirus transmission to the fetus. *J Med Virol* (2013) 85(11):1960–7. doi:10.1002/jmv.23691
71. Brizic I, Susak B, Arapovic M, Huszthy PC, Hirsl L, Kvestak D, et al. Brain-resident memory CD8(+) T cells induced by congenital CMV infection prevent brain pathology and virus reactivation. *Eur J Immunol* (2018). doi:10.1002/eji.201847526
72. Marcoe JP, Lim JR, Schaubert KL, Fodil-Cornu N, Matka M, McCubbrey AL, et al. TGF- β is responsible for NK cell immaturity during ontogeny and increased susceptibility to infection during mouse infancy. *Nat Immunol* (2012) 13(9):843–50. doi:10.1038/ni.2388
73. Wu X, Chen Y, Wei H, Sun R, Tian Z. Development of murine hepatic NK cells during ontogeny: comparison with spleen NK cells. *Clin Dev Immunol* (2012) 2012:759765. doi:10.1155/2012/759765

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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